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b. Reference Number : SS21-078 Review  
c. SS21-078 Trackcanges AB.doc
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6. Paper accepted (12 Juli 2024)
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1. Submission received for Makara Journal of Science  
( 21 Desember 2021)



Denai Wahyuni &lt;denaiwahyuni69@htp.ac.id&gt;

**MS #1303: Submission received for Makara Journal of Science****Editors of Makara Journal of Science** <editors-science-1303@dcuischolarhub.bepress.com>

21 Desember 2021 pukul 14.15

Kepada: =?UTF-8?Q?=22denaiwahyuni69=40htp=2Eac=2Eid=22?= &lt;denaiwahyuni69@htp.ac.id&gt;

Cc: The Authors &lt;authors-science-1303@dcuischolarhub.bepress.com&gt;, The Editors &lt;editors-science-1303@dcuischolarhub.bepress.com&gt;

**A new submission** for Makara Journal of Science has been uploaded by "[denaiwahyuni69@htp.ac.id](mailto:denaiwahyuni69@htp.ac.id)" <[denaiwahyuni69@htp.ac.id](mailto:denaiwahyuni69@htp.ac.id)>.

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The title is:

"VEGETABLE INSECTICIDE EXTRACT OF SCENTED ROOT (Polygala paniculata)  
IN CONTROLLING Aedes aegypti MOSQUITO"

The keywords are:

extract P. paniculata, Aedes aegypti mosquito, vegetable insecticide

The disciplines are:

Life Sciences | Organisms | Public Health

The submission has been assigned #1303. Please refer to this number in any correspondence related to the submission.

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Best regards,

The Editors

Makara Journal of Science

## 2. Revision of Manuscript (27 Mei 2022)

- Dokumen:
- a. Reference Number : SS21-078
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Denai Wahyuni &lt;denaiwahyuni69@htp.ac.id&gt;

**SS21-078 Revision of Manuscript**

**Makara Journal of Science** <editor\_mss@ui.ac.id>  
Kepada: Denai Wahyuni <denaiwahyuni69@htp.ac.id>  
Cc: "Dr. Ivandini Tribidasari A." <ivandini.tri@sci.ui.ac.id>

27 Mei 2022 pukul 13.47

Dear Author,

We would like to inform you that your article titled "Vegetable Insecticide Extract of Scented Root (*Polygala paniculata*) in Controlling *Aedes aegypti* Mosquito" (SS21-078) has been reviewed by reviewers.

Based on the comments from two reviewers, this manuscript will be acceptable with major revision. Please indicate the changes that have been made with point-by-point responses to the reviewer's comments (response letter). If you are unable to revise according to the reviewer's recommendation, unfortunately, your manuscript can not be published in our journal.


Please respond to the comments of reviewers and submit a revised manuscript within 15 days of this notification. Thank you for submitting your manuscript to Makara Journal of Science.

Sincerely yours,

Chief Editor  
Makara Journal of Science  
Prof. Dr. Ivandini Tribidasari A.

**4 lampiran**

 **SS21-078 AB.rtf**  
108K

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82K

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475K



**Response letter.doc**

34K

## **2.a. KOREKSI.... SS21-078 AB**

### **Referee's Report**

**Reference Number : SS21-078**

**Title : Vegetable Insecticide Extract of Scented Root (*Polygala paniculata*) in Controlling *Aedes aegypti* Mosquito**

Please provide an overall assessment of the paper and provide answers to the following questions. Detailed comments and recommendations should not be written in the text, but should be numerically listed on separate sheets of paper that do not reveal your identity or affiliation. Minor wording or grammatical changes can be indicated in red pen or pencil in the text.

**A. Please give your *appreciation of the scientific interest and novelty* of results described**  
(in Indonesia or in English)

**This manuscript has novelty since Scented Root (*Polygala paniculata*) which has potential as an insecticide has never been tested on *Aedes aegypti* mosquitoes.**

**Unfortunately, this paper is written in poor English and does not meet the rules of academic English. many expressions and translations that do not conform to academic rules are found in this manuscript. the author should use a language consultant whenever this manuscript is accepted.**

**B. Style and Organization** (Please check as appropriate)

**Double Click**

- |  |   |
|--|---|
| 1. Is it clearly presented, well organized, and clearly written?   | <input checked="" type="checkbox"/> Yes <input type="checkbox"/> No |
| 2. Does it contain superfluous material?   | <input type="checkbox"/> Yes <input checked="" type="checkbox"/> No |
| 3. Is the title appropriate?   | <input checked="" type="checkbox"/> Yes <input type="checkbox"/> No |
| 4. Does the abstract include the important points of the paper   | <input checked="" type="checkbox"/> Yes <input type="checkbox"/> No |
| 5. If applicable, is the experiment section sufficiently detailed?   | <input type="checkbox"/> Yes <input checked="" type="checkbox"/> No |
| 6. Is sufficient information included or cited to support the assertions made and conclusions drawn?   | <input checked="" type="checkbox"/> Yes <input type="checkbox"/> No |
| 7. Are references to related work adequate, up to date and readily available?  | <input checked="" type="checkbox"/> Yes <input type="checkbox"/> No |
| 8. Are the illustrations and tables all necessary and adequate?  | <input type="checkbox"/> Yes <input checked="" type="checkbox"/> No |
| 9. Are the figure and table captions complete and accurate?  | <input checked="" type="checkbox"/> Yes <input type="checkbox"/> No |
| 10. Are the conclusions satisfactory, sound and justified  | <input checked="" type="checkbox"/> Yes <input type="checkbox"/> No |
| 11. Do you agree with the "Prime Novelty" as indicated (by the author)?<br>Why or why not? It is first time try on <i>Aedes aegypti</i><br>..... | <input checked="" type="checkbox"/> Yes <input type="checkbox"/> No |
| 12. Does the manuscript contain original and self-consistent ideas?<br>Please comment .....  | <input checked="" type="checkbox"/> Yes <input type="checkbox"/> No |
| 13. Does the manuscript contain subject matter that might/should be omitted?<br>If so, what? .....   | <input type="checkbox"/> Yes <input checked="" type="checkbox"/> No |







## 2.b. KOREKSI 2.....SS21-078 AMR

### Referee's Report

Reference Number : SS21-078 Review

Title : **Vegetable Insecticide Extract of Scented Root (*Polygala paniculata*) in Controlling *Aedes aegypti* Mosquito**

Please provide an overall assessment of the paper and provide answers to the following questions. Detailed comments and recommendations should not be written in the text, but should be numerically listed on separate sheets of paper that do not reveal your identity or affiliation. Minor wording or grammatical changes can be indicated in red pen or pencil in the text.

**A. Please give your appreciation of the scientific interest and novelty of results described**  
(in Indonesia or in English)

The manuscript presents an investigation of the insecticidal effect of *Polygala paniculata* for the control of *Aedes aegypti*. It is an original and relevant work for research with insecticides obtained from natural products, however, the work should include more statistical analysis to accurately estimate the time needed to kill 50% of mosquitoes.

**B. Style and Organization** (Please check as appropriate)

**Double Click**

- |     |   |   |
|-----|---|---|
| 0   | Is it clearly presented, well organized, and clearly written?                                     | <input type="checkbox"/> Yes <input checked="" type="checkbox"/> No |
| 1   | Does it contain superfluous material?   | <input type="checkbox"/> Yes <input checked="" type="checkbox"/> No |
| 2   | Is the title appropriate?   | <input checked="" type="checkbox"/> Yes <input type="checkbox"/> No |
| 3   | Does the abstract include the important points of the paper                                       | <input checked="" type="checkbox"/> Yes <input type="checkbox"/> No |
| 4   | If applicable, is the experiment section sufficiently detailed?                                   | <input checked="" type="checkbox"/> Yes <input type="checkbox"/> No |
| 5   | Is sufficient information included or cited to support the assertions made and conclusions drawn? | <input checked="" type="checkbox"/> Yes <input type="checkbox"/> No |
| 7.  | Are references to related work adequate, up to date and readily available?                        | <input checked="" type="checkbox"/> Yes <input type="checkbox"/> No |
| 8.  | Are the illustrations and tables all necessary and adequate?                                      | <input checked="" type="checkbox"/> Yes <input type="checkbox"/> No |
| 9.  | Are the figure and table captions complete and accurate?  | <input type="checkbox"/> Yes <input checked="" type="checkbox"/> No |
| 10. | Are the conclusions satisfactory, sound and justified   | <input checked="" type="checkbox"/> Yes <input type="checkbox"/> No |
| 11. | Do you agree with the "Prime Novelty" as indicated (by the author)?<br>Why or why not? .....      | <input checked="" type="checkbox"/> Yes <input type="checkbox"/> No |
| 12. | Does the manuscript contain original and self-consistent ideas?<br>Please comment .....           | <input checked="" type="checkbox"/> Yes <input type="checkbox"/> No |
| 13. | Does the manuscript contain subject matter that might/should be omitted?<br>If so, what? .....    | <input type="checkbox"/> Yes <input checked="" type="checkbox"/> No |
| 14. | For manuscript in English, Is the English satisfactory?   | <input type="checkbox"/> Yes <input checked="" type="checkbox"/> No |

**C. Quality and Assessment**

(Please check as appropriate)

- Makes major contributions to the advancement of the subject
- Sound, original, and of interest
- Sound, original, but not suitable for Makara Seri Sains
- Does not add to knowledge of the subject
- Contains fundamental errors or faulty judgements

**D. Referee's Recommendation**

E. (Please check all boxes that apply)

- Acceptable in present form
- Acceptable with grammatical revision
- Acceptable with minor technical revision
- Acceptable with major technical revision
- Submit to further refereeing :  
(Suggest name of referee) .....
- Reject

Check here if you want to see the Revised Version

Check here if you made annotation on the manuscript

**E. Timeliness of Research**

New

Somewhat New

Old

**F. Scientific Impact**

High

Moderate

No Impact

**G. Match between title, abstract, data and conclusions**

Excellent

Good

Poor

**H. Rate scientific novelty to the community**

High

Medium

Low

**I. Rate the interest to the community**

High

Medium

Low

**J. Remarks/Additional Comments (if any)**

(in Indonesia or in English)

1. Consider using *Ae. aegypti* as an abbreviation for *Aedes aegypti*.

2. The abbreviations YFV, ZIKV, CHIKV and DENV are used to refer to viruses (etiological agents) and not to diseases. Consider tweaking your text (page 01)

3. There is redundancy between the Materials and Methods topic presented in the subtopics "Preparation of Materials and Tools" and "Extract Making". The text needs to be adjusted to avoid such redundancies (page 2-3).

4. In Figure 1, only one caption should be kept for an image. The descriptions that are in the figure must be placed in the legend (page 4).

5. When presenting the data in Figure 01, it is interesting to consider the average number of deaths by concentration and not the sum of deaths (Line 135).

6. Note the writing of the scientific name "*Aedes aegypti*" on pages 5 and 6.

7. In table 01 the date presented in the table (0.000) are different from those discussed in the text (0.001) (line

167).

**8.** The value 0.08 is greater than 0.05, not less. (line 185)

**9.** The Time required for 50% knockdown of mosquitoes ( $KT_{50}$ ) could be accurately estimated using probit analysis (table 4).

**10.** In obtaining table 05, were the mortality averages considered?

**11.** In line 196 refers to table 04 and in line 207 refers to table 05.

**12.** According to the presentation of the text, I suggest that the Results section be presented separately from the Discussion section.

1 **2.c. SS21-078 Trackhanges AB (2)**

2  
3  
4  
5  
6

**Vegetable Insecticide Extract of Scented Root (*Polygala paniculata*) in Controlling *Aedes aegypti* Mosquito**

**Comment [AB1]:** Please revise the title with appropriate english, vegetable insecticide is not comon to use in this title

7 **ABSTRACT**

8 Controlling *Aedes aegypti* mosquitoes with chemical insecticides causes resistance effects on  
9 humans, environmental residues, as well as contaminates food and water. *Polygala paniculata*  
10 has potential as an alternative of insecticide in controlling *A. aegypti* mosquitoes. This study  
11 further aimed to determine the effect of the Insecticide Score of *P. paniculata* extract on  
12 *A. aegypti* mosquitoes' mortality based on  $KT_{50}$ . The 20 mosquitoes were used to examine the  
13 effectiveness of 10%, 15%, 20%, 25% of *P. paniculata* concentration with positive and negative  
14 controls in four repetitions every five minutes for one hour. The results showed the effect of  
15 extract *P. paniculata* on *A. aegypti* mosquitoes' mortality. The Kruskal-Wallis test resulted in a p-  
16 value of  $0.001 < 0.05$ , while the Spearman Correlation test gave a p-value of  $0.008 < 0.05$ .  
17 Furthermore, the correlation strength was + 0.312 or 31.2%, with 10% concentration, while 15%  
18 had  $KT_{50}$  with Insecticide Scores of 1 and 2, implying no knockdown effect. A 20%  
19 concentration had  $KT_{50}$  with an Insecticide Score of 3, indicating a weak knockdown effect,  
20 while a 25% concentration had  $KT_{50}$  with an Insecticide Score of 5, signifying a quick  
21 knockdown effect. This shows that a 25% concentration has a quick knockdown time in *A.*  
22 *aegypti* mosquitoes. Therefore, extract *P. paniculata* may have the potential as a vegetable  
23 insecticide in controlling *A. aegypti* mosquitoes.

**Comment [AB2]:** It is wrong way to abbreviate Aedes. Please follow the rule!!!

24 **Keywords:** extract *P. paniculata*, *Aedes aegypti* mosquito, vegetable insecticide

25 **1. Introduction**

26 The *Aedes aegypti* mosquito (L) belongs to the Order Diptera and family Culicidae, the  
27 main vector often neglected as a transmitter of diseases in humans. The diseases include yellow  
28 fever (YFV), Zika virus (ZIKV), Chikungunya (CHIKV), dengue virus (DENV), and other  
29 arboviruses [1]–[4]. The human arboviral disease transmitted by *A. aegypti* is a global public  
30 health threat [5]. This disease causes significant morbidity and mortality in developing countries  
31 [2]. The transmission of dengue fever is increasing in urban and semi-urban areas in tropical  
32 countries worldwide [6]. It is estimated that 40% or 50–528 million people worldwide are at risk  
33 of becoming infected with dengue fever and around 10,000–20,000 people die yearly [7].  
34 According to WHO, about 390 million cases of dengue virus infection occur every year, of  
35 which 96 million manifest clinically with high severity. Furthermore, WHO reports that about  
36 3.9 billion people are at risk of being infected with the dengue virus [8].

37 Indonesia is one of the developing countries where dengue hemorrhagic fever (DHF) is an  
38 infectious disease and an unresolved health problem. In 2020, there were 15,132 DHF cases in  
39 Indonesia, with a death rate of 145 people and a DHF incident rate of 31.23 [9]. This implies  
40 problems in efforts to control the disease.

**Comment [AB3]:** DHF not only problem in developing conuntry. Please rewrite the paragraph

41 There is no specific vaccine for dengue fever, hence its control is conducted by eradicating  
42 the mosquito vector using synthetic insecticides [2]–[4], [10], [11]. However, long-term  
43 application and extensive use of synthetic insecticides cause the accumulation of residues in

**Comment [AB4]:** It is not relate between vaccine, insecticide, vector. Author can delete this paragraph. Focus, please...

44 food, water, soil and give adverse health effects on humans and ecosystems [12], [13]. This  
45 leaves residues that pollute the environment [4], [14] increasing population resistance of *A.*  
46 *aegypti* [3], [5], [15]–[19].

47 Controlling *A. aegypti* mosquitoes using synthetic insecticides involves fumigation with  
48 pyrethroids and larvacides containing temephos [11]. Pyrethroid resistance causes *A. aegypti*  
49 to keep growing, increasing the incidence of dengue fever (DF) worldwide. This increases the risk  
50 of almost half of the world's population being infected with the disease [5]. Similarly, using  
51 temephos to kill larvae increases the resistance of mosquitoes larvae, *A. aegypti*. This has been  
52 reported in several countries, such as Brazil [3], Mexico [20] and Southeast Asia countries,  
53 including Indonesia [5], Malaysia, Philippines, Thailand, Singapore, Laos, and Myanmar [21]

54 There is a need to obtain alternative insecticides effective in controlling the mosquito  
55 population *A. aegypti* [22]. This could be achieved using natural plant chemical compounds with  
56 the potential to control the mosquito population effectively. The environmentally friendly  
57 method would guarantee plant protection and avoid the side effects of synthetic insecticides [13],  
58 [23], [24]. Bioactive plant compounds are biodegradable, environmentally friendly, and non-  
59 toxic to other insects [1]. This is because bio-insecticides only affect the target insect without  
60 destroying beneficial natural enemies. Additionally, they are a safe, economical, target-specific,  
61 biodegradable, and residue-free environmental food source [23], [25].

62 *Polygala paniculata*, also known as vetiver in Riau, Indonesia, is a good-smelling annual  
63 herbaceous plant belonging to the family Polygalaceae and the genus Polygala. The plant is often  
64 used as traditional medicine, a tonic, and in inflammation cases of asthma, bronchitis, arthritis,  
65 and other pathologies, as well as kidney disorders [26]. Moreover, *P. paniculata* is used for in-  
66 vivo protection against the neurotoxic effects of Methylmercury (Hg) [27]. bronchitis,  
67 neurahenia, inflammation, amnesia, topical anesthetic, and expectorant drugs [28]. The Polygala  
68 extract produces secondary metabolites, including alkaloids, saponins, flavonoids, phenols,  
69 tannins, steroids, and terpenoids [28]. Similarly, *P. paniculata* contains alkaloids, flavonoids,  
70 tannins, saponins, and steroids [29]. Therefore, these bioactive compounds make *P. paniculata*  
71 potentially useful as a vegetable insecticide. This becomes an interesting study topic because the  
72 bioactive compounds extracted from *P. paniculata* have never been explored as vegetable  
73 insecticides in controlling the *A. aegypti* mosquito. Therefore, it is important to examine the  
74 insecticide score of the toxin contained in the bioactive compounds of *P. paniculata* in killing the  
75 *A. aegypti* mosquito.

## 77 2. Materials and Methods

### 79 Preparation of Materials and Tools

80 This study used 2500 grams of plant *P. paniculata* obtained in Kubang Region Siak Hulu  
81 sub-district, Kampar, Riau. This plant was proven by a laboratory certification test at the  
82 Botanical Laboratory, Faculty of Mathematics and Natural Sciences, Riau University. The test  
83 used 96% ethanol to macerate the *P. paniculata* plant powder. Furthermore, distilled water was  
84 used as a solvent and as a negative control in the extraction process, while synthetic insecticide  
85 Baygon cypermethrin was used as a positive control. The *A. aegypti* mosquito was obtained from  
86 breeding kept at the Parasitology Laboratory of Abdurrah University Pekanbaru. The equipment  
87 used in this study includes a blender, analytical scales, Rotary Vacuum Evaporator, stopwatch,  
88 shaker water bath, thermometer, hygrometer, Buchner funnel, stick, basin, test box, syringe, and  
89 a spray bottle.

**Comment [AB5]:** Explain the distribution of this plant, in the world and in Indonesia.

## 90 **Test Animal Preparation**

91 Test animals were bred using media containing clean water in a cool place and protected  
92 from direct sunlight for the *A. aegypti* mosquitoes to lay their eggs. The larvae were reared in an  
93 aquarium at 24.2<sup>0</sup>C - 24.4<sup>0</sup>C, with a relative humidity of 67-70%. The larvae were fed with  
94 coconut water [30] to become larvae instar III and IV before becoming mosquitoes. The adult  
95 *A. aegypti* mosquitoes were then used as test animals.

## 96 **Extract Making**

97 2500 grams of *P. paniculata* plants were washed and air-dried at room temperature and  
98 blended them to form 400 grams of powder. The powder was macerated with 96% ethanol until  
99 completely submerged for three days. The resulting solution was filtered using a Buchner funnel  
100 and placed in a dark bottle. Furthermore, the dregs from the first filtering were then soaked again  
101 for one day, filtered, and the process repeated in the third immersion. The results from the three  
102 maceration processes were combined and concentrated using a Vacuum Rotary Evaporator to  
103 evaporate 96% ethanol and obtain an extract. The extract obtained was stored in the refrigerator  
104 to be used later [31].

Comment [AB6]: Use good english please

## 105 **Testing**

106 The test was conducted by transferring 20 *A. aegypti* mosquitoes to each test boxes and  
107 spraying them with various *P. paniculata* plant extract, K (+), and K(-) concentrations four  
108 times. The effect of the concentrations on the *A. aegypti* mosquito was observed by looking at  
109 the changes in behavior, movement, and physical condition until death. The dead of *A. aegypti*  
110 mosquitoes were counted every five minutes for one hour. Moreover, the insecticide score of  
111 *P. paniculata* was determined from the number of *A. aegypti* mosquitoes considered dead  
112 at 5-minute intervals. At the end of each treatment, *A. aegypti* mosquitoes that remained alive  
113 were left to die or killed with Baygon.

Comment [AB7]: Explain the kind of testing

## 114 **Data analysis**

115 Data were analyzed using the Statistical Test of Analysis of Variance with RAL, followed  
116 by the One Way ANOVA test. However, when the One Way ANOVA test does not meet the  
117 requirements, the Kruskal-Wallis Non-Parametric Test and the Test Spearman Analysis are  
118 conducted to examine the relationship between the independent and dependent variables.

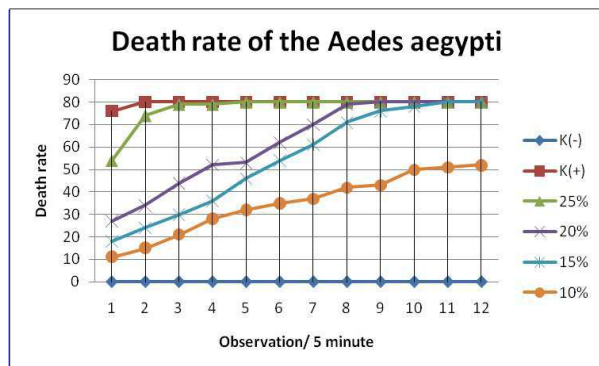
## 119 **3. Results and Discussion**

### 122 **The results of observing the *A. aegypti* mosquitoes' death after spraying the *P. paniculata* 123 **plant extract****

124 The *A. aegypti* mosquitoes died after spraying the *P. paniculata* plant extract at 10% and  
125 15% concentrations. They died slowly by flying irregularly and actively and falling in a tilted  
126 body position. The mosquitoes appeared weak, with some legs still moving, before becoming  
127 paralyzed, dying, and their bodies remaining stiff. The death was faster at 20% and 25%  
128 concentrations. In the positive control (K+), death occurred in less than five minutes, while in the  
129 negative control (K-), the *A. aegypti* mosquitoes tried to avoid the spray during the four  
130 experiment repetitions.

### 131 **Total mortality of *A. aegypti* mosquitoes at each concentration with four repetitions**

133



Comment [AB8]: Aedes aegypti italic

134

135  
136

Figure 1. Total mortality of *A. aegypti* mosquitoes at each concentration with four repetitions (Primary Data. 2020)

137 Figure 1 shows that 10%, 15%, 20%, and 25% concentrations of the *P. paniculata* plant  
 138 extract were sprayed to the treatment group. In the first five minutes of observation, the four  
 139 repetitions of spraying caused the mortality of 11, 18, 27, and 54 *A. aegypti* mosquitoes. The  
 140 mortality increased to 52, 80, 80, and 80 in the 60<sup>th</sup> minute. In positive control (K+) using  
 141 synthetic insecticide Baygon (cypermethrin), the four repetitions of spraying caused the death of  
 142 76 *A. aegypti* mosquitoes in less than five minutes. The test in the negative control (K-) using  
 143 distilled water did not cause death. The highest total mortality of 80 *A. aegypti* mosquitoes at  
 144 15% concentration occurred at 51-55 minutes. Furthermore, a 20% concentration caused 80  
 145 deaths at 41-45 minutes, while a 25% concentration caused 80 deaths at 21-25 minutes.

Comment [AB9]: Use good English please

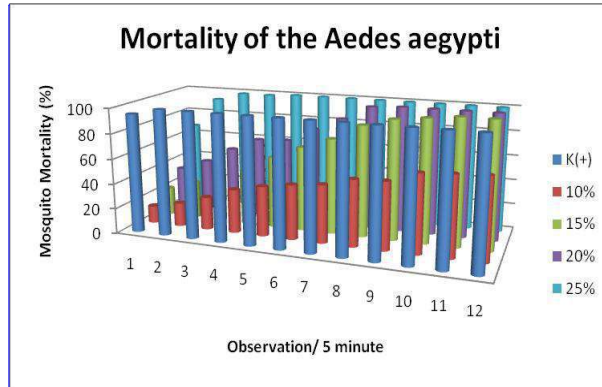
146

147 **The death percentage of *A. aegypti* mosquitoes at each concentration after every five**  
 148 **minutes of observation**

149 In Figure 2, the treatment group was sprayed with 10%, 15%, 20%, and 25%  
 150 concentrations of the *P. paniculata* plant extract. The mortality percentage of *A. aegypti*  
 151 mosquitoes in four repetitions was 13.75%, 22.5%, 33.75%, and 67.5% in five minutes of the  
 152 observation. The total mortality increased to 65%, 100%, 100%, and 100% in the 60<sup>th</sup> minute.  
 153 The positive control (K+) test using synthetic insecticide Baygon (cypermethrin) caused a 95%  
 154 mortality, where the first *A. aegypti* mosquito died in less than five minutes during the four test  
 155 repetitions. In contrast, the negative control (K-) test using distilled water did not cause death.  
 156 The highest total mortality of 100% *A. aegypti* mosquitoes at 15% concentration occurred at 51-  
 157 55 minutes. The 20% and 25% concentrations caused 80 deaths at 41-45 and 21-25 minutes,  
 158 respectively.

159





**Comment [AB10]:** Not clear, please make in 2 dimension graph

160

Figure 2: Percentage of *A. aegypti* mosquito mortality at each concentration every five minutes of observation (Primary Data, 2020)

161

162

163

**Kruskal Wallis Non-Parametric Test**

**a. Concentration**

164

165

166

167

Table 1. Non-Parametric Test Results Kruskal Wallis Concentration

	Knockdown
Chi-Square	30.324
Df	5
Asymp. Sig.	.000

**Comment [AB11]:** No need to put the statistical analysis in here. Write as result please

168

Primary Data, 2020

169

170

171

172

173

174

175

176

**b. Time**

Table 2. Non-Parametric Test Results Kruskal Wallis (Time)

	Knockdown
Chi-Square	19.512
Df	11
Asymp. Sig.	.052

177

Primary Data, 2020

178

179

180

181

In the Non-Parametric Kruskal Wallis, the significant value is  $0.052 > 0.05$ . This implies a significant difference in the *A. aegypti* mosquito mortality between the treatment groups with different mean rank values. Therefore, the contact time of *P. paniculata* plant extracts

**Comment [AB12]:** No need to put the statistical analysis in here. Write as result please

182 significantly affects the mortality of *A. aegypti* mosquitoes because the significant value is >  
 183 0.05.

184  
 185 **Spearman Analysis**

186 **Table 3. Spearman Analysis Test Results**

	Concentration	Time	Knockdown
Spearman's rho			
	Concentration	Koefisien korelasi	1.000
		Sig. (2-tailed)	.000
		N	72
			72

187 Primary Data, 2020

**Comment [AB13]:** No need to put the statistical analysis in here. Write as result please

188 The Spearman Analysis test results show a significant value of 0.08 <0.05. This indicates a  
 189 correlation between the different concentrations of *P. paniculata* plant extracts and the mortality  
 190 of *A. aegypti* mosquitoes. The correlation strength is denoted by the coefficient of 0.312 or  
 191 31.2%. Moreover, the positive correlation means that the higher concentration of *P. paniculata*  
 192 plant extract increased the number of deaths of *A. aegypti* mosquitoes.

193 **Insecticide Score of *P. paniculata* Plants**

194 Table 4. The average percentage of death of *A. aegypti* mosquitoes at various  
 195 concentrations of treatment with Knockdown Time (KT<sub>50</sub>)

196

Time	10%	15%	20%	25%	K (+)	K (-)
5	2,75	4,5	6,75	13,5	19	0
10	3,75	6	8,5	18,5	20	0
15	5,25	7,5	11	19,75	20	0
20	7	9	13	19,75	20	0
25	8	11,5	13,25	20	20	0
30	8,75	13,5	15,5	20	20	0
35	9,25	15,25	17,5	20	20	0
40	10,5	17,75	19,75	20	20	0
45	10,75	19	20	20	20	0
50	12,5	19,5	20	20	20	0
55	12,75	20	20	20	20	0
60	13	20	20	20	20	0

197 Primary Data, 2020

**Comment [AB14]:** Please make the table more clear. Explain the highlighting data!

198 In Table 1, the test repetition using a 10% concentration of the *P. paniculata* plant extract  
 199 produced KT<sub>50</sub> between 36-40 minutes. The test using 15%, 20%, and 25% concentrations  
 200 produced KT<sub>50</sub> between 21-25, 11-15, and less than five minutes, respectively. On average, no *A.*  
 201 *aegypti* mosquitoes fell in the negative control, while KT<sub>50</sub> occurred in less than five minutes in  
 202 the positive control. This means that the different concentrations of *P. paniculata* plant extracts  
 203 affected the number of *A. aegypti* differently during each treatment and repetition.  
 204

205  
206  
207

Table 5: Insecticide Score of *P. paniculata* Plant Extract Based on  $KT_{50}$

Group Control	KT50 (Time)	Knockdown Effect	Insecticide Score	Interpretation
Concentration 10%	36-40	-	1	-
Concentration 15%	21-25	-	2	-
Concentration 20%	11-15	+	3	Weak Knockdown
Concentration 25%	< 5	+++	5	Quick Knockdown
Positive Control	< 5	+++	5	Quick Knockdown

Primary Data, 2020

Comment [AB15]: What mean +?

208  
209

210 Table 2 shows the effectiveness level of the four *P. paniculata* plant extract  
 211 concentrations and positive control based on Insecticide Knockdown Time<sub>50</sub> (KT<sub>50</sub>). A 10%  
 212 concentration of the *P. paniculata* plant extract had a KT of<sub>50</sub> between 36-40 minutes. This  
 213 implies an Insecticide Score of 1 or no knockdown effect. Similarly, a 15% concentration of the  
 214 plant extracts had a KT of<sub>50</sub> between 21-25 minutes, indicating an Insecticide Score of 2 or no  
 215 knockdown. A 20% concentration of the plant extract had a KT of<sub>50</sub> between 11-15 minutes,  
 216 implying an Insecticide Score of 3 or a weak knockdown effect. Furthermore, a 25%  
 217 concentration of *P. paniculata* plant extract had a KT<sub>50</sub> in less than 5 minutes. This indicates an  
 218 Insecticide Score of 5 or a quick knockdown effect. The 25% concentration had a KT<sub>50</sub> in less  
 219 than 5 minutes, similar to the positive control, with an Insecticide Score of 5 or a quick  
 220 knockdown effect. Therefore, the 25% concentration of *P. paniculata* plant extract was most  
 221 effective in knocking out *A. aegypti* mosquitoes, with an Insecticide Score of 5 or a quick  
 222 knockdown effect.

223 The results in Figures 1 and 2 show that the number and percentage of *A. aegypti*  
 224 mosquito deaths increased with the concentration of *P. paniculata* plant extract. Higher  
 225 concentrations increase the accumulation of *P. paniculata* plant extract's toxic bioactive  
 226 compounds in *A. aegypti* mosquitoes' bodies, increasing their mortality.

227 Kosini examined the effect of the *Gnidia kaussiana* (*Thymeleaceae*) extract on  
 228 *Callosobruchus maculatus*. The study explained that increased absorption of toxic compounds in  
 229 the *Gnidia kaussiana* extract accelerates the mortality process of larvae *Callosobruchus*  
 230 *maculatus* by melanizing the cuticle. This disrupts the endocrine system due to the presence of  
 231 secondary metabolites such as terpenoids, alkaloids, and flavonoids [32]. Furthermore, another  
 232 study examined the effect of *Ocimum basilicum*, vegetable insecticides, on the death of *A.*  
 233 *aegypti* mosquitoes. The results showed that the mortality of *A. aegypti* mosquitoes increases  
 234 with an increase in the absorption of toxic compounds in the *O. basilicum* extract. Also, longer  
 235 exposure to the extract increases the toxicity [33]. A previous study examined the effect of  
 236 vegetable larvicides of *Carbera manghas* leaves on *A. aegypti* mosquito larvae. The findings  
 237 showed the larvicide toxicity increased with the *C. manghas* extract's concentration and  
 238 exposure time. This is because the absorption of more toxic compounds affects the body's  
 239 metabolism and increases the mortality of *A. aegypti* larvae [34].

240 Many previous studies stated that the bioactive plant compounds have insecticidal,  
 241 larvicidal, repellent, and environmentally-friendly effects useful for insect control. According to  
 242 Hikal, essential oils, flavonoids, alkaloids, glycosides, esters, and fatty acids have anti-insect  
 243 effects. Therefore, they could be used as an alternative to chemical compounds in insect control

244 as repellents, feeding deterrents or antifeedants, toxicants, inhibitors, growth factors,  
245 chemosterilants, and attractants [23].

246 Previous studies explained that *Polygala* plants contain bioactive compounds with  
247 various biological activities, such as alkaloids, saponins, flavonoids, phenols, tannins, steroids,  
248 and terpenoids [28], [29]. Tannins and flavonoids are phenolic plant compounds that act as  
249 primary antioxidants or free radical scavengers [35]. The bioactive compounds of *P. paniculata*  
250 able to affect the mortality of *A. aegypti* mosquitoes are flavonoids, saponins, tannins, alkaloids,  
251 steroids, and terpenoids. In this study, the *A. aegypti* mosquitoes death was caused by  
252 compounds in the *P. paniculata* plant extract. The compounds entered the mosquito's body  
253 through contact or respiratory poisoning and the mouth and digestive tract, causing stomach  
254 poisoning.

255 The analysis shows that the number of *A. aegypti* mosquitoes that died when exposed to  
256 the *P. paniculata* plant extract varied with the extract's concentration. Variations in the  
257 concentrations affected the mortality of *A. aegypti* mosquitoes differently in each treatment and  
258 repetition. The results in Figures 1 and 2 show that spraying a 25% concentration of the *P.*  
259 *paniculata* plant extract for 60 minutes kills 80 *A. aegypti* mosquitoes in less than the first five  
260 minutes. This is the largest number of deaths that occurred faster than other concentrations.  
261 Therefore, the higher concentration of the plant extract increased its effectiveness as a vegetable  
262 insecticide against mosquitoes.

263 This study compared the effectiveness level of the four *P. paniculata* plant extract  
264 concentrations with positive controls based on Insecticide Knockdown Time<sub>50</sub> (KT<sub>50</sub>). A 25%  
265 concentration of the plant extract had a KT<sub>50</sub> of less than five minutes, implying an Insecticide  
266 Score of 5 or a quick knockdown effect. This is in line with the 2006 WHO standard, which  
267 stated that an insecticide is has a knockdown time required to drop a vector when the median  
268 knockdown ranges between 3-5. Furthermore, it has a quick knockdown effect when it has a  
269 KT<sub>50</sub> of less than five minutes. According to Norris, a good insecticide requirement for  
270 controlling disease vector insect species must cause a rapid knockdown of the target species,  
271 especially active pathogens. Additionally, it must quickly intervene and kill adult mosquitoes  
272 [16].

273 The 25% concentration had a KT<sub>50</sub> of less than five minutes, similar to a positive control  
274 Baygon containing cypermethrin. They both had an Insecticide Score 5, implying a quick  
275 knockdown effect. Therefore, the 25% concentration of the *P. paniculata* plant extract was the  
276 most effective in knocking down *A. aegypti* mosquitoes. This is because it had an Insecticide  
277 Score of 5 or a quick knockdown effect. The positive control treatment was intended to compare  
278 the quality of *P. paniculata* plant extract concentration. In contrast, the negative control  
279 treatment was used to compare its effectiveness with the plant extract. The results showed that no  
280 mosquito died after 60 minutes of observation.

281 Chang highlighted the need to use vegetable insecticides as an alternative insect  
282 controller. Using inappropriate insecticides causes insects to adapt easily by metabolic  
283 detoxification quickly and survival. This causes synchronization of insect immunity passed on to  
284 their offspring. Therefore, it is necessary to develop safe alternative insecticides, larvicides, and  
285 repellents effective for humans, animals, the environment, and the ecosystem. Natural  
286 insecticides are needed to suppress vector resistance and slow down their genetic adaptation [36].  
287 According to Hikal et al., botanical insecticides only affect target insects, do not destroy  
288 beneficial natural enemies, and are a safe and residue-free food source. Hikal et al. recommended  
289 using plant-based insecticides as an integrated insect management program that greatly reduces

290 the use of synthetic insecticides [23]. It is more environmentally friendly, effective, cheap, and  
291 naturally available [14].  
292

### 293 **Conclusion**

294 The active substance contained in the *P. paniculata* plant extract has the ability to drop  
295 and kill *A. aegypti* mosquitoes. This is because more compounds in the plant extracts exposed to  
296 the mosquitoes increase the knockdown effect time. Therefore, the plant extract is a potential  
297 alternative insecticide for controlling *A. aegypti* mosquitoes. It does not leave residues in the  
298 environment and is safe for other living beings. Also, the extract's compounds do not cause  
299 resistance against *A. aegypti* mosquitoes, and the plant has economic value and is beneficial to  
300 cultivate.

### 301 **Acknowledgments**

302 This study was funded by DIPA STIKes Hang Tuah Pekanbaru in 2020 with the title  
303 "Insecticide of Fragrant Root Extract (*Polygala paniculata*) in Control of Mosquitoes *Aedes*  
304 *aegypti*," with Contract Number No. 08/STIKes-HTP/VI/2020/0147, A.

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**Comment [AB16]:** 1.Please change the scopus non-indexed reference to the scopus indexed reference

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3. First revision: received with major revision (24-Juni-2022)

- Dokumen:
- a. Response Letter Reviewer 1
  - b. Response Letter Reviewer 2
  - c. Revised Result Mayor



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## SS21-078 Revision of Manuscript

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**Makara Journal of Science** <editor\_mss@ui.ac.id>  
Kepada: Denai Wahyuni <denaiwahyuni69@htp.ac.id>  
Cc: "Dr. Ivandini Tribidasari A." <ivandini.tri@sci.ui.ac.id>

24 Juni 2022 pukul 10.07

Dear Author,

Your revision has been received by us nicely and needs to be checked from our editor.  
Please kindly resubmit the revision and response letter to the website.  
For the cooperation, we say thank you.

Best Regards,

Puji Astuti  
Editorial Assistant  
Makara Journal of Science

[Kutipan teks disembunyikan]

### 3. a. Response letter Reviewer 1

Journal : Makara Journal of Science

Manuscript ID :

Title: **Bio-insecticides Extract of Scented Root (*Polygala paniculata*) in Controlling *Aedes aegypti* Mosquito**

There are some following issues that needs to be addressed:

Reviewer 1

<b>Reviewer's Comments</b>	<b>Author's Comments</b>
<b>Reference Number : SS21-078;</b> Comment (AB1). Please revise the title with appropriate english, vegetable insecticide is not comon to use in this title	Thank you, we have corrected the suggestion regarding the title, here are the improvements:  Bio-insecticide's Extract of Scented Root ( <i>Polygala paniculata</i> ) in Controlling <i>Aedes aegypti</i> Mosquito
Comment (AB2): It is wrong way to abbreviate Aedes. Please follow the rule!!!	Thank you for the suggestion regarding the correct abbreviation regarding <i>Aedes</i> to <i>Ae. aegypti</i> we have corrected everything in the manuscript regarding the title we have corrected, here are the corrections:
Comment (AB3): DHF not only problem in developing conutry. Please rewrite the paragraph	Thank you for your input regarding this paragraph, we have corrected it in the text on lines 34-37 as follows:  Indonesia is one of where dengue hemorrhagic fever (DHF) is an infectious disease and an unresolved health problem. In 2020, there were 15,132 DHF cases in Indonesia, with a death rate of 145 people and a DHF incident rate of 31.23[9].. This implies problems in efforts to control the disease.
Comment (AB4): It is not relate between vaccine, insecticide, vector. Author can delete this paragraph. Focus, please...	Thank you for your suggestion regarding there is no relationship between vaccines , insecticides, vectors we have corrected in the text on lines 38-41 as follows:  However, long-term application and extensive use of synthetic insecticides cause the accumulation of residues in food, water, soil and give adverse health effects on humans and ecosystems[10], [11]. This leaves residues that pollute the environment [3], [12]. increasing population resistance of <i>Ae. aegypti</i> [4], [5], [13]–[17].
Comment (AB5): Explain the dsitribution this plant, in the world and in Indonesia	Thank you for your suggestion to explain the distribution of this plant in the world and Indonesia, we have added it to the text on lines 57-63 as follows:

	<p><i>Polygala L.</i> is one of the largest genera belonging to the <i>Polygalaceae</i> tribe. This genus consists of 500 species and can be found in tropical, sub-tropical, temperate and mountainous areas throughout the world except New Zealand. Most of these species grow in Central and South Tropical America. Several types of <i>Polygala L.</i> that can be used as medicine such as :<i>Polygala chinensis L.</i>, <i>Polygala paniculata L.</i>, <i>Polygala polifoliaPresl.</i>, and <i>Polygala sibirica L.</i>[24].<i>Polygala paniculata</i>, also known as vetiver in Riau, Indonesia, is a good-smelling annual herbaceous plant belonging to the family Polygalaceae and the genus Polygala.</p>
<p>Comment (AB6): Use good english please</p>	<p>Thank you for We have corrected the correction regarding the subtitles to use good English in the text on line 91 as follows</p> <p>Extraction Process</p>
<p>Comment (AB7): Explain the kind of testing</p>	<p>Thank you for the correction regarding the type of test we have corrected and added to the text on line 100-109 as follows:</p> <p><b>Bio-insecticide’s Extract of <i>P. paniculata</i> Test against <i>Ae. Aegypti</i> Mosquitos</b></p> <p>Bio-insecticide’s extract of <i>P. paniculata</i> tests against <i>Ae. aegypti</i> was carried out by transferring 20 <i>Ae. aegypti</i> in each test boxes, then sprayed with various concentrationsof <i>P. paniculata</i> plant extract with four repetitions, as well as for K (+) and K(-).The effect of the concentrations on the <i>A. aegypti</i> mosquito was observed by looking at the changes in behavior, movement, and physical condition until death. <i>Ae. Aegypti</i> mosquitoes death were counted every five minutes for one hour. Moreover, the insecticide score of <i>P. paniculata</i> was determined from the number of <i>Ae. Aegypti</i> mosquitoes considered deadat 5-minute intervals. <i>Ae. Aegypti</i> mosquitoes that remained alive were left to die or killed with Baygon.</p>
<p>Comment (AB9): Use good English please</p>	<p>Thank you for the correction to use correct English, we have corrected the text on line 134-143 as follows:</p> <p>Based on Figure 1 In the treatment group, the <i>P. paniculata</i> plant extract was sprayed with 10%, 15%, 20% and 25% concentration in the first 5 minutes of observation. <i>Ae.aegypti</i> mosquitoes death rate with 4 repetitions in a row was 11, 18, 27 and 54 individuals. In the 60 minutes, the total mortality of <i>Ae. aegypti</i> mosquitoes in 4 repetitions was 52, 80, 80 and 80. In positive control (K+) using synthetic insecticide baygon (cypermethrin) in less than 5 minutes <i>Ae. aegypti</i> mosquitoes fell and died in 4 repetitions, namely 76 individuals.While in the negative control (+) using distilled water did not cause death.There was 80 <i>Ae. aegypti</i></p>

	<p>mosquitoes were death at a concentration of 15% occurred at 51-55 minutes, a 20% concentration of 80 total deaths occurred at 41-45 minutes and a 25% concentration of 80 total deaths occurred at 21-25 minutes.</p>
<p>Comment (AB10): Not clear, please make in 2 dimension graph</p>	<p>Thank you for the correction regarding the three (3) dimensional graph we have corrected to a two (2) dimensional graph in the text on lines 162-168 as follows:</p> <div data-bbox="635 526 1497 1019" data-label="Figure"> <p>The figure is a grouped bar chart with 12 groups of bars on the x-axis, labeled 'Observation/ 5 minutes' from 1 to 12. The y-axis is 'Death rate Ae. Aegypti (%)' ranging from 0 to 120. Each group contains six bars representing different concentrations: K(-) (blue), K(+) (red), 10% (green), 15% (purple), 20% (teal), and 25% (orange). The death rate generally increases over the 12 observations and with higher concentrations. For example, at observation 1, the death rates are approximately 10% for K(-), 20% for K(+), 30% for 10%, 40% for 15%, 50% for 20%, and 60% for 25%. By observation 12, all concentrations reach a death rate of 100%.</p> </div> <p>Figure 2: The Average Mortality of <i>Ae. aegypti</i> at each Concentration of Five Minutes Observation</p>
<p>Comment (AB11); (AB12); (AB13): No need to put the statistical analysis in here. Write as result please</p>	<p>Thank you for the correction to not include statistical analysis, we have changed it to include only the results in the text on lines 169-188 as follows:</p> <p>Based on the results of study in figure 1 and 2, explained that the different concentrations of <i>P. paniculata</i> plant extracts gave different effects on number of <i>Ae. aegypti</i> mosquitoes death in each treatment and repetition. The number of <i>Ae.aegypti</i> mosquitoes death tends to increase along with the increase in the concentration of <i>P. paniculata</i> plant extracts. It's means that the higher used of the concentration, the higher of potency of the <i>P. paniculata</i> plant extract as a bioinsecticide against the <i>Ae. Aegypti</i> mosquito. During one hour observations showed the increasing number of <i>Ae. Aegypti</i> mosquitos's death. It's explained the longer an observation time, the greater potential as a bioinsecticide. This is supported by the results of the Kruskal-Wallis test (p-value of 0.001 &lt;0.05), which means that there is a significant difference between the death rate of the <i>Ae. aegypti</i> mosquito and the difference in the concentration of the <i>P. paniculata</i> plant extract (10%, 15%, 20%, 25 %) of Knockdown Time acceleration. The results of the Kruskal-Wallis test in this study were correct and continued with the Spearman correlation test with the results of p-value 0.008 &lt;0.05 there was a significant correlation between the increase in the concentration</p>

of the extract (treatment group) and the knockdown time of the *P. paniculata* plant extract. From the results of the Kruskal-Wallis test and the Spearman correlation test, it can be concluded that the greater of concentration, the faster the time of death of *Ae. aegypti* mosquito, the strength of the correlation is denoted by a correlation coefficient 0.312 (31.2%). Correlation coefficient occurred with positive value means that correlation is moderate. The higher concentrations of *P. paniculata* plants extract then the faster of knockdown time

Comment (AB14): Please make the table more clear. Explain the highlighting data!

Thanks for the suggestion we have corrected the explanation on lines 194-198 in the manuscript

Table 1 Average of *Ae. Aegypti* Falling Down At Various Treatment Concentrations With Knockdown Time<sub>50</sub> (KT<sub>50</sub>)

Time	10%	15%	20%	25%	K (+)	K (-)
5	2,75	4,5	6,75	13,5	19	0
10	3,75	6	8,5	18,5	20	0
15	5,25	7,5	11	19,75	20	0
20	7	9	13	19,75	20	0
25	8	11,5	13,25	20	20	0
30	8,75	13,5	15,5	20	20	0
35	9,25	15,25	17,5	20	20	0
40	10,5	17,75	19,75	20	20	0
45	10,75	19	20	20	20	0
50	12,5	19,5	20	20	20	0
55	12,75	20	20	20	20	0
60	13	20	20	20	20	0

: *Ae. Aegypti* Knockdown Time(KT<sub>50</sub>);

Comment (AB15) untuk tabel 5. What mean +?

Thank you we tried to explain Regarding the meaning of + In table 5 : Insecticide Score of *P. paniculata* Plant Extract Based on KT<sub>50</sub>.

The knockdown effect has been defined by WHO 2006 in the book Guidelines for Testing Mosquitos Adulticides for Indoor Residual Spraying and Treatment of Mosquitos Nets That: Insecticide Score Based on KT 50

KT50 (minute)	Score	Knockdown effect	Interpretation
>50	0	-	-
31-49	1	-	-
16-30	2	-	-
11-15	3	+	Weak Knockdown
5-10	4	++	Strong Knockdown
<5	5	+++	Quick Knockdown

source: WHO, 2006

A value of 3 means the median knockdown is in the range of 11-15 minutes which is interpreted to have a knockdown

	<p>effect (+) but is weak. A value of 4 means the median knockdown is in the range of 5-10 minutes which is interpreted to have a strong knockdown effect (++). A value of 5 means the median knockdown is in the range of less than 5 minutes which is interpreted to have a knockdown effect (+++) that the insecticide has a “Quick Knockdown Effect” (WHO, 2006).</p> <p>So the + sign means explaining the Knockdown effect status of Bioinsecticide</p>
<p>Comment (AB16): Please change the scopus non-indexed reference to the scopus indexed reference</p>	<p>Thank you for the correction to change all non-Scopus references to Scopus-indexed references, all of which we have changed to Scopus-indexed references in the script as follows</p> <p><b>References</b></p> <p>[1] L. H. Guimarães de Oliveira <i>et al.</i>, “Agave sisalana extract induces cell death in <i>Aedes aegypti</i> hemocytes increasing nitric oxide production,” <i>Asian Pac. J. Trop. Biomed.</i>, vol. 6, no. 5, pp. 396–399, May 2016.</p> <p>[2] D. A. Fernandes <i>et al.</i>, “Larvicidal compounds extracted from <i>helicteres velutina</i> K. Schum (Sterculiaceae) evaluated against <i>aedes aegypti</i> L.,” <i>Molecules</i>, vol. 24, no. 12, p. 2315, 2019.</p> <p>[3] L. Scalvenzi <i>et al.</i>, “Larvicidal activity of <i>Ocimum campechianum</i>, <i>Ocotea quixos</i> and <i>Piper aduncum</i> essential oils against <i>Aedes aegypti</i>,” <i>Parasite</i>, vol. 26, no. 23, pp. 1–8, 2019.</p> <p>[4] D. Valle, D. F. Bellinato, P. F. Viana-Medeiros, J. B. P. Lima, and A. D. J. Martins Junior, “Resistance to temephos and deltamethrin in <i>aedes aegypti</i> from Brazil between 1985 and 2017,” <i>Mem. Inst. Oswaldo Cruz</i>, vol. 114, no. 3, pp. 1–17, Mar. 2019.</p> <p>[5] Z. H. Amelia-yap, C. D. Chen, M. Sofian-azirun, and V. L. Low, “Pyrethroid resistance in the dengue vector <i>Aedes aegypti</i> in Southeast Asia : present situation and prospects for management,” <i>Parasit. Vectors</i>, vol. 11, no. 332, pp. 1–17, 2018.</p> <p>[6] S. Leta, T. Jibat, E. M. De Clercq, K. Amenu, M. U. G. Kraemer, and C. W. Revie, “International Journal of Infectious Diseases Global risk mapping for major diseases transmitted by <i>Aedes aegypti</i> and <i>Aedes albopictus</i>,” <i>Int. J. Infect. Dis.</i>, vol. 67, pp. 25–35, 2018.</p> <p>[7] M. Y. Lee, “Essential Oils as Repellents against</p>

	<p>Arthropods,” <i>Biomed Res. Int.</i>, vol. -, no. -, pp. 1–9, 2018.</p> <p>[8] WHO, “Dengue and severe dengue,” <i>World Health Organization</i>, 19-May-2021. [Online]. Available: <a href="https://www.who.int/news-room/fact-sheets/detail/dengue-and-severe-dengue">https://www.who.int/news-room/fact-sheets/detail/dengue-and-severe-dengue</a>. [Accessed: 06-Dec-2021].</p> <p>[9] Kemenkes RI, <i>Incidence Rate Per 100.000 Penduduk Demam Berdarah Dengue Tahun 2010-2018</i>. Jakarta, 2020.</p> <p>[10] K. Baana, H. Angwech, and G. M. Malinga, “Ethnobotanical survey of plants used as repellents against housefly, <i>Musca domestica</i> L. (Diptera: Muscidae) in Budondo Subcounty, Jinja District, Uganda,” <i>J. Ethnobiol. Ethnomed.</i>, vol. 14, no. 1, pp. 1–8, May 2018.</p> <p>[11] A. T. H. Mossa, S. M. M. Mohafrash, and N. Chandrasekaran, “Safety of natural insecticides: Toxic effects on experimental animals,” <i>Biomed Res. Int.</i>, vol. 2018, pp. 1–18, 2018.</p> <p>[12] E. A. Rohmah, S. Subekti, and M. Rudyanto, “Larvicidal Activity and Histopathological Effect of <i>Averrhoa bilimbi</i> Fruit Extract on <i>Aedes aegypti</i> from Surabaya, Indonesia,” <i>J. Parasitol. Res.</i>, vol. 2020, pp. 1–5, Aug. 2020.</p> <p>[13] C. S. Bibbs, D. A. Hahn, P. E. Kaufman, and R. Xue, “Sublethal effects of a vapour-active pyrethroid, transfluthrin, on <i>Aedes aegypti</i> and <i>Ae. albopictus</i> (Diptera: Culicidae) fecundity and oviposition behaviour,” <i>Parasit. Vectors</i>, vol. 11, no. 1, p. 486, Dec. 2018.</p> <p>[14] P. A. Rojas-Pinzón, . Juan, J. Silva-Fernández, and J. Dussán, “Laboratory and simulated-field bioassays for assessing mixed cultures of <i>Lysinibacillus sphaericus</i> against <i>Aedes aegypti</i> (Diptera: Culicidae) larvae resistant to temephos,” <i>Applied Entomol. Zool.</i>, vol. 53, pp. 183–191, 2018.</p> <p>[15] E. L. R. De Sá <i>et al.</i>, “Evaluation of insecticide resistance in <i>aedes aegypti</i> populations connected by roads and rivers: The case of tocantins state in Brazil,” <i>Mem. Inst. Oswaldo Cruz</i>, vol. 114, no. 2, pp. 1–10, Feb. 2019.</p> <p>[16] S. Murfadunnisa <i>et al.</i>, “Larvicidal and enzyme inhibition of essential oil from <i>Spheranthus amaranthoides</i> (Burm.) against lepidopteran pest <i>Spodoptera litura</i> (Fab.) and their impact on non-target earthworms,” <i>Biocatal. Agric.</i></p>
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	<p><i>Biotechnol.</i>, vol. 21, no. 101324, pp. 1–9, 2019.</p> <p>[17] E. J. Norris, A. D. Gross, L. C. Bartholomay, and J.R. Coats, “Plant essential oils synergize various pyrethroid insecticides and antagonize malathion in <i>Aedes aegypti</i>,” <i>Med. Vet. Entomol.</i>, vol. 33, pp. 453–466, 2019.</p> <p>[18] J. G. Galvão <i>et al.</i>, “<i>Lippia gracilis</i> essential oil in <math>\beta</math>-cyclodextrin inclusion complexes: an environmentally safe formulation to control <i>Aedes aegypti</i> larvae,” <i>Pest Manag. Sci.</i>, vol. 75, no. 2, pp. 452–459, Feb. 2019.</p> <p>[19] Y. Kandel <i>et al.</i>, “Widespread insecticide resistance in <i>Aedes aegypti</i> L. From New Mexico, U.S.A.,” <i>PLoS One</i>, vol. 14, no. 2, pp. 1–16, Feb. 2019.</p> <p>[20] G. N. Pandiyan, N. Mathew, and S. Munusamy, “Larvicidal activity of selected essential oil in synergized combinations against <i>Aedes aegypti</i>,” <i>Ecotoxicol. Environ. Saf.</i>, vol. 174, pp. 549–556, 2019.</p> <p>[21] J. K. Suluvoy and V. M. Berlin Grace, “Phytochemical profile and free radical nitric oxide (NO) scavenging activity of <i>Averrhoa bilimbi</i> L. frui extract,” <i>3 Biotech</i>, vol. 7, no. 1, pp. 1–11, May 2017.</p> <p>[22] H. D. Manh, D. T. Hue, N. Thi, T. Hieu, D. Thi, and T. Tuyen, “The Mosquito Larvicidal Activity of Essential Oils from <i>Cymbopogon</i> and <i>Eucalyptus</i> Species in Vietnam,” <i>MPDI</i>, vol. 11, no. 128, pp. 1–7, 2020.</p> <p>[23] B. Sisay, T. Tefera, M. Wakgari, G. Ayalew, and E. Mendesil, “The efficacy of selected synthetic insecticides and botanicals against fall armyworm, <i>spodoptera frugiperda</i>, in maize,” <i>Insects</i>, vol. 10, no. 2, pp. 1–14, Feb. 2019.</p> <p>[24] J. L. C. . Van Valkenburg, <i>Plant Resources of south-east asia 12 (2) Medicinal and poisonous plant 2</i>. Bogor, Indonesia: Bogor Prosea, 2002.</p> <p>[25] F. da R. Lapa <i>et al.</i>, “Antinociceptive Properties of the Hydroalcoholic Extract and the Flavonoid Rutin Obtained from <i>Polygala paniculata</i> L. in Mice,” <i>Basic Clin. Pharmacol. Toxicol.</i>, vol. 104, no. 4, pp. 306–315, Apr. 2009.</p> <p>[26] M. Farina <i>et al.</i>, “Protective effects of <i>Polygala paniculata</i> extract against methylmercury- induced neurotoxicity in</p>
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	<p>mice,” <i>J. Pharm. Pharmacol.</i>, pp. 1503–1508, 2005.</p> <p>[27] J. L. C. Da Rocha, R. F. A. De Tripodi Calumby, D. F. Da Silva, H. N. Brandão, C. F. Villarreal, and F. O. De Lima, “Evaluation of biological activity of polygala boliviensis in experimental models,” <i>Iran. J. Pharm. Res.</i>, vol. 18, no. 2, pp. 793–802, Mar. 2019.</p> <p>[28] J. D. Fontana <i>et al.</i>, “Accelerating the Morphogenetic Cycle of the Viral Vector <i>Aedes aegypti</i> Larvae for Faster Larvicidal Bioassays,” <i>Biomed Res. Int.</i>, vol. 2020, 2020.</p> <p>[29] Kemenkes RI, “Farmakope Herbal Indonesia,” II., Jakarta, 2017.</p> <p>[30] WHO, <i>Mosquito Adulticides for Indoor Residual Spraying and Treatment of Mosquito Nets</i>. Geneva: World Health Organization, 2006.</p> <p>[31] D. Kosini and E. N. Nukenine, “Bioactivity of novel botanical insecticide from gnidia kaussiana (Thymeleaceae) against callosobruchus maculatus (Coleoptera: Chrysomelidae) in stored vigna subterranea (Fabaceae) grains,” <i>J. Insect Sci.</i>, vol. 17, no. 1, pp. 1–7, 2017.</p> <p>[32] G. S. da Botas <i>et al.</i>, “molecules Baccharis reticularia DC. and Limonene Nanoemulsions: Promising Larvicidal Agents for <i>Aedes aegypti</i> (Diptera: Culicidae) Control,” <i>mdpi.com</i>, vol. 22, no. -, pp. 2–14, 2017.</p> <p>[33] I. Sina, Zaharah, and M. S. M. Sabri, “Larvicidal activities of extract flower averrhoa bilimbi L. Towards important species mosquito, anopheles barbirostris (diptera: Culicidae),” <i>Int. J. Zool. Res.</i>, vol. 12, no. 1–2, pp. 25–31, 2016.</p> <p>[34] X. Chang <i>et al.</i>, “Multiple Resistances and Complex Mechanisms of <i>Anopheles sinensis</i> Mosquito: A Major Obstacle to Mosquito-Borne Diseases Control and Elimination in China,” <i>PLoS Negl. Trop. Dis.</i>, vol. 8, no. 5, p. e2889, May 2014.</p>
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### **3.b. Response letter Reviewer 2**

<b>Reviewer's Comments</b>	<b>Author's Comments</b>
1. Consider using <i>Ae. aegypti</i> as an abbreviation for <i>Aedes aegypti</i> .	Thank you for providing corrections to change the abbreviation <i>Aedes aegypti</i> to <i>Ae. Aegypti</i> , we have corrected this suggestion in the manuscript
2. The abbreviations YFV, ZIKV, CHIKV and DENV are used to refer to viruses (etiological agents) and not to diseases. Consider tweaking your text (page 01)	<p>Thank you for the suggestion to consider changing the text regarding the abbreviations YHF, ZIKV, CHIKV and DENV in paragraph 1. We have changed this in the text on lines 23-25 as follows:</p> <p>The <i>Aedes aegypti</i> mosquito (L) belongs to the Order Diptera and family Culicidae, the main vector often neglected as a transmitter of diseases in humans. The diseases include Yellow Fever, Zica, Chikungunya, Dengue Haemorrhagic Fever and other arbo viruses[1]–[4].</p>
3. There is redundancy between the Materials and Methods topic presented in the subtopics "Preparation of Materials and Tools" and "Extract Making". The text needs to be adjusted to avoid such redundancies (page 2-3).	<p>Thank you for the correction</p> <p>Regarding the redundancy between the topic of Materials and Methods presented in the subtopic Preparation of Materials and Tools and Preparation of Extracts in the text, we have corrected it on lines 78-83 in the text as follows:</p> <p><b>Preparation of Materials and Tools</b></p> <p>This study used 2500 gram <i>P. paniculata</i>, 5 litre of 96% ethanol, 5 litre of distilled water, 1 bottle of synthetic insecticide Baygon (cypermethrin), and 480 <i>Ae. aegypti</i> mosquito. The equipment used in this study includes a blender, analytical scales, Rotary Vacuum Evaporator, stopwatch, shaker water bath, thermometer, hygrometer, Buchner funnel, stick, basin, test box, syringe, and a spray bottle.</p>
4. In Figure 1, only one caption should be kept for an image. The descriptions that are in the figure must be placed in the legend (page 4).	Thank you for the correction regarding the information in Figure 1, we have corrected it on line 144-147 as follows:

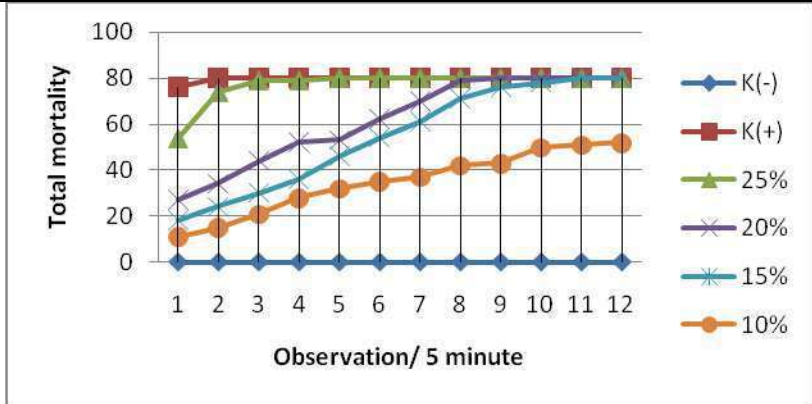


Figure 1 *Ae. Aegypti* mosquitoes total mortality at each concentration with four repetitions

5. When presenting the data in Figure 01, it is interesting to consider the average number of deaths by concentration and not the sum of deaths (Line 135).

Thank you for the advice. Figure 01 is retained but the average mortality is made in Figure 02 as follows: on lines 144-147 and lines 164-168

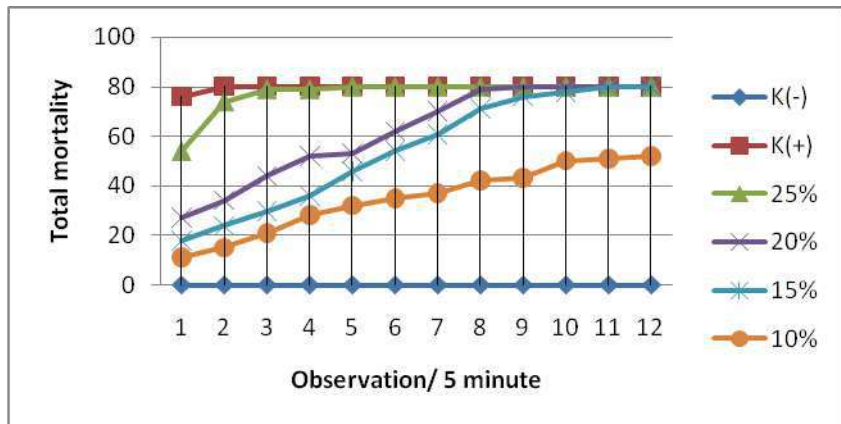


Figure 1 *Ae. Aegypti* mosquitoes total mortality at each concentration with four repetitions

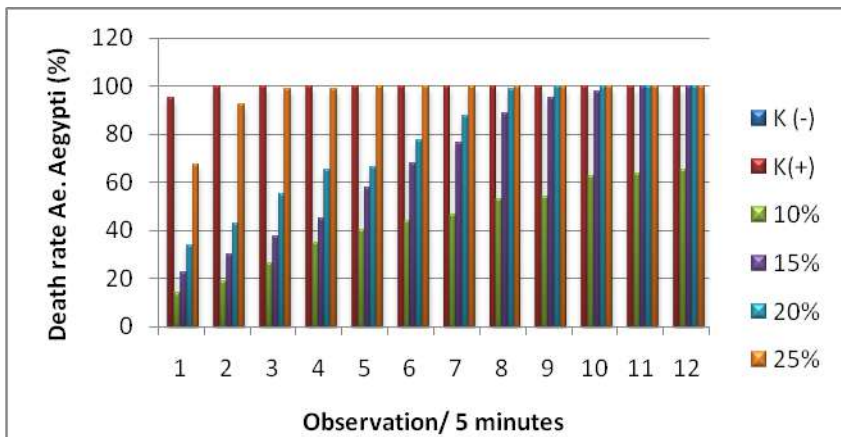


	Figure 2: The Average Mortality of <i>Ae. aegypti</i> at each Concentration of Five Minutes Observation
6. Note the writing of the scientific name " <i>Aedes aegypti</i> " on pages 5 and 6.	Thank you for the advice regarding scientific writing <i>Aedes aegypti</i> " on pages 5 and 6 we have corrected it as suggested
7. In table 01 the date presented in the table (0.000) are different from those discussed in the text (0.001) (line 167)	<p>Thank you for the advice. Based on input from other reviewers that this table is not needed but the results from the table. This is conveyed in the script narrative only on lines 169-188 as follows:</p> <p>Based on the results of study in figure 1 and 2, explained that the different concentrations of <i>P. paniculata</i> plant extracts gave different effects on number of <i>Ae. aegypti</i> mosquitoes death in each treatment and repetition. The number of <i>Ae.aegypti</i> mosquitoes death tends to increase along with the increase in the concentration of <i>P. paniculata</i> plant extracts. It's means that the higher used of the concentration, the higher of potency of the <i>P. paniculata</i> plant extract as a bioinsecticide against the <i>Ae. Aegypti</i> mosquito. During one hour observations showed the increasing number of <i>Ae. Aegypti</i> mosquitos's death. It's explained the longer an observation time, the greater potential as a bioinsecticide. This is supported by the results of the Kruskal-Wallis test (p-value of <math>0.001 &lt; 0.05</math>), which means that there is a significant difference between the death rate of the <i>Ae. aegypti</i> mosquito and the difference in the concentration of the <i>P. paniculata</i> plant extract (10%, 15%, 20%, 25 %) of Knockdown Time acceleration. The results of the Kruskal-Wallis test in this study were correct and continued with the Spearman correlation test with the results of p-value <math>0.008 &lt; 0.05</math> there was a significant correlation between the increase in the concentration of the extract (treatment group) and the knockdown time of the <i>P. paniculata</i> plant extract. From the results of the Kruskal-Wallis test and the Spearman correlation test, it can be concluded that the greater of concentration, the faster the time of death of <i>Ae. aegypti</i> mosquito, the strength of the correlation is denoted by a correlation coefficient 0.312 (31.2%). Correlation coefficient occurred with positive value means that correlation is moderate. The higher concentrations of <i>P. paniculata</i> plants extract then the faster of knockdown time</p>
8. The value 0.08 is greater than 0.05, not less. (line 185)	Thanks for the correction. We have deleted this information because the related table has been changed to a new narrative
9. The Time required	Thank you for the suggestion, to use probit analysis to determine the

<p>for 50% knockdown of mosquitoes (KT50) could be accurately estimated using probit analysis (table 4).</p>	<p>time required for 50% mosquito knockdown (KT50) in Table 4  We try to explain  Knockdown time (KT50) means the time it takes for each concentration of <i>P. paniculata</i> Bioinsecticide to drop 50% of the average <i>Ae. aegypti</i>. We did not use the probit analysis in Table 4, because here we only looked at how many minutes the average number of mosquitoes fell by 50% for each concentration of <i>P. paniculata</i> Bioinsecticide.</p>
<p>10. In obtaining table 05, were the mortality averages considered?</p>	<p>Thank you for the correction.  We try to explain,  We have changed Table 05 to Table 02 because Tables 01, 02, 03 have been omitted according to input from other reviewers. For Table 02, we do not use the average mortality, but we use <math>KT_{50}</math> (Time) which is the time it takes to drop 50% of <i>Ae. aegypti</i> after being sprayed with <i>P. paniculata</i> Bioinsecticide at each test concentration.</p>
<p>11. In line 196 refers to table 04 and in line 207 refers to table 05.</p>	<p>Thank you for the correction. We have corrected the script on line 201-202 as follows:    Knockdown Time<sub>50</sub> (<math>KT_{50}</math>) is the times to knockdown of <i>Ae. aegypti</i> mosquitoes. In Table 1, the average repetition of the study show during the 10% concentration of <i>P. paniculata</i> plant</p>
<p>12. According to the presentation of the text, I suggest that the Results section be presented separately from the Discussion section.</p>	<p>Thank you for the advice. We have corrected the manuscript according to the following suggestions:    A. Results pada line 117  B. Discussion pada line 229</p>

### **3.c. Revised result mayor**

#### **Bio-insecticide's Extract of Scented Root (*Polygala paniculata*) in Controlling *Aedes aegypti* Mosquito**

#### **ABSTRACT**

Controlling *Aedes aegypti* mosquitoes with chemical insecticides causes resistance effects on humans, environmental residues, as well as contaminates food and water. *Polygala paniculata* has potential as an alternative of insecticide in controlling *Ae. Aegypti* mosquitoes. This study further aimed to determine the effect of the Insecticide Score of *P. paniculata* extract on *Ae. Aegypti* mosquitoes' mortality based on  $KT_{50}$ . The 20 mosquitoes were used to examine the effectiveness of 10%, 15%, 20%, 25% of *P. paniculata* concentration with positive and negative controls in four repetitions every five minutes for one hour. The results showed the effect of extract *P. paniculata* on *Aeaegypti* mosquitoes' mortality. The Kruskal-Wallis test resulted in a p-value of  $0.001 < 0.05$ , while the Spearman Correlation test gave a p-value of  $0.008 < 0.05$ . Furthermore, the correlation strength was + 0.312 or 31.2%, with 10% concentration, while 15% had  $KT_{50}$  with Insecticide Scores of 1 and 2, implying no knockdown effect. A 20% concentration had  $KT_{50}$  with an Insecticide Score of 3, indicating a weak knockdown effect, while a 25% concentration had  $KT_{50}$  with an Insecticide Score of 5, signifying a quick knockdown effect. This shows that a 25% concentration has a quick knockdown time in *Ae. Aegypti* mosquitoes. Therefore, extract *P. paniculata* may have the potential as a bio-insecticides in controlling *Ae. aegypti* mosquitoes.

Keywords: extract *P. paniculata*, *Aedes aegypti* mosquito, bio-insecticides

#### **1. Introduction**

The *Aedes aegypti* mosquito (L) belongs to the Order Diptera and family Culicidae, the main vector often neglected as a transmitter of diseases in humans. The diseases include yellow fever (YFV), Zika virus (ZIKV), Chikungunya (CHIKV), dengue virus (DENV), and other arboviruses[1]–[4]. The human arboviral disease transmitted by *Ae.aegypti* is a global public health threat[5]. This disease causes significant morbidity and mortality in developing countries [1]. The transmission of dengue fever is increasing in urban and semi-urban areas in tropical countries worldwide[6].. It is estimated that 40% or 50–528 million people worldwide are at risk of becoming infected with dengue fever and around 10,000–20,000 people die yearly[7]. According to WHO, about 390 million cases of dengue virus infection occur every year, of which 96 million manifest clinically with high severity. Furthermore, WHO reports that about 3.9 billion people are at risk of being infected with the dengue virus[8]

Indonesia is one of where dengue hemorrhagic fever (DHF) is an infectious disease and an unresolved health problem. In 2020, there were 15,132 DHF cases in Indonesia, with a death rate of 145 people and a DHF incident rate of 31.23[9].. This implies problems in efforts to control the disease. However, long-term application and extensive use of synthetic insecticides cause the accumulation of residues in food, water, soil and give adverse health effects on humans and ecosystems[10], [11]. This leaves residues that pollute the environment [3], [12]. increasing population resistance of *Ae. aegypti*[4], [5], [13]–[17].



43 Controlling *Ae.aegypti* mosquitoes using synthetic insecticides involves fumigation with  
44 pyrethroids and larvacides containing temephos[18].Pyrethroid resistance causes *Ae. aegypti* to  
45 keep growing, increasing the incidence of dengue fever (DF) worldwide. This increases the risk  
46 of almost half of the world's population being infected with the disease[5]. Similarly, using  
47 temephos to kill larvae increases the resistance of mosquitoes larvae, *A. aegypti*. This has been  
48 reported in several countries, such as Brazil[4], Mexico[19]and Southeast Asia countries,  
49 including Indonesia Malaysia, Philippines, Thailand, Singapore, Laos, and Myanmar[5].

50 There is a need to obtain alternative insecticides effective in controlling the mosquito  
51 population *A. aegypti*[20]. This could be achieved using natural plant chemical compounds with  
52 the potential to control the mosquito population effectively. The environmentally friendly  
53 method would guarantee plant protection and avoid the side effects of synthetic insecticides,[10],  
54 [21], [22].Bioactive plant compounds are biodegradable, environmentally friendly, and non-toxic  
55 to other insects[2].This is because bio-insecticides only affect the target insect without  
56 destroying beneficial natural enemies. Additionally, they are a safe, economical, target-specific,  
57 biodegradable, and residue-free environmental food source[23].

58 *Polygala L.* is one of the largest genera belonging to the *Polygalaceae* tribe. This genus  
59 consists of 500 species and can be found in tropical, sub-tropical, temperate and mountainous  
60 areas throughout the world except New Zealand. Most of these species grow in Central and  
61 South Tropical America. Several types of *Polygala L.* that can be used as medicine such as  
62 *:Polygala chinensis L., Polygala paniculata L., Polygala polifoliaPresl., and Polygala sibirica*  
63 *L.*[24].*Polygala paniculata*, also known as vetiver in Riau, Indonesia, is a good-smelling annual  
64 herbaceous plant belonging to the family Polygalaceae and the genus Polygala. The plant is often  
65 used as traditional medicine, a tonic, and in inflammation cases of asthma, bronchitis, arthritis,  
66 and other pathologies, as well as kidney disorders[25]. Moreover, *P. paniculata* is used for in-  
67 vivo protection against the neurotoxic effects of Methylmercury (Hg)[26], bronchitis,  
68 neurahenia, inflammation, amnesia, topical anesthetic, and expectorant drugs[27]. The Polygala  
69 extract produces secondary metabolites, including alkaloids, saponins, flavonoids, phenols,  
70 tannins, steroids, and terpenoids[27].Therefore, these bioactive compounds make *P. paniculata*  
71 potentially useful as a bio-insecticides. This becomes an interesting study topic because the  
72 bioactive compounds extracted from *P. paniculata* have never been explored as bio-insecticides  
73 in controlling the *Ae. aegypti* mosquito. Therefore, it is important to examine the insecticide  
74 score of the toxin contained in the bioactive compounds of *P. paniculata* in killing the *Ae.*  
75 *aegypti* mosquito.

76

## 77 **2. Materials and Methods**

78

### 79 **Preparation of Materials and Tools**

80 This study used 2500 gram softplant *P. paniculata* obtained in Kubang Region Siak Hulu  
81 sub-district, Kampar, Riau. This plant was proven by a laboratory certification test at the  
82 Botanical Laboratory, Faculty of Mathematics and Natural Sciences, Riau University. The test  
83 used 96% ethanol to macerate the *P. paniculata* plant powder. Furthermore, distilled water was  
84 used as a solvent and as a negative control in the extraction process, while synthetic insecticide  
85 Baygon (cypermethrin) was used as a positive control. The *A. aegypti* mosquito was obtained  
86 from breeding kept at the Parasitology Laboratory of Abdurrah University Pekanbaru. The  
87 equipment used in this study includes a blender, analytical scales, Rotary Vacuum Evaporator,

88 stopwatch, shaker water bath, thermometer, hygrometer, Buchner funnel, stick, basin, test box,  
89 syringe, and a spray bottle.

### 90 **Test Animal Preparation**

91 Test animals were bred using media containing clean water in a cool place and protected  
92 from direct sunlight for the *Ae. Aegypti* mosquitoes to lay their eggs. The larvae were reared in  
93 an aquarium at 24.2<sup>0</sup>C - 24.4<sup>0</sup>C, with a relative humidity of 67-70%. The larvae were fed with  
94 coconut water[28] to become larvae instar III and IV before becoming mosquitoes. The adult  
95 *Ae.aegypti* mosquitoes were then used as test animals.

### 96 **Extraction Process**

97 2500 grams of *P. paniculata* plants were washed and air-dried at room temperature and  
98 blended them to form 400 grams of powder. The powder was macerated with 96% ethanol until  
99 completely submerged for three days. The resulting solution was filtered using a Buchner funnel  
100 and placed in a dark bottle. Furthermore, the dregs from the first filtering were then soaked again  
101 for one day, filtered, and the process repeated in the third immersion. The results from the three  
102 maceration processes were combined and concentrated using a Vacuum Rotary Evaporator to  
103 evaporate 96% ethanol and obtain an extract. The extract obtained was stored in the refrigerator  
104 to be used later[29].

### 105 **Bio-insecticide's Extract of *P. paniculata* Test against *Ae. Aegypti* Mosquitos**

106  
107 Bio-insecticide's extract of *P. paniculata* tests against *Ae. aegypti* was carried out by  
108 transferring 20 *Ae. aegypti* in each test boxes, then sprayed with various concentrations of *P.*  
109 *paniculata* plant extract with four repetitions, as well as for K (+) and K(-).The effect of the  
110 concentrations on the *A. aegypti* mosquito was observed by looking at the changes in behavior,  
111 movement, and physical condition until death. *Ae. Aegypti* mosquitoes death were counted every  
112 five minutes for one hour. Moreover, the insecticide score of *P. paniculata* was determined from  
113 the number of *Ae. Aegypti* mosquitoes considered dead at 5-minute intervals. *Ae. Aegypti*  
114 mosquitoes that remained alive were left to die or killed with Baygon.

### 115 **Data analysis**

116 Data were analyzed using the Statistical Test of Analysis of Variance with RAL, followed  
117 by the One Way ANOVA test. However, when the One Way ANOVA test does not meet the  
118 requirements, the Kruskal-Wallis Non-Parametric Test and the Test Spearman Analysis are  
119 conducted to examine the relationship between the independent and dependent variables.

120

## 121 **3. Results and Discussion**

### 122 **A. Results**

123

#### 124 **The Results of Observing *Ae.aegypti* Mosquitoes' Death After Spraying the *P. paniculata*** 125 **Plant Extract**

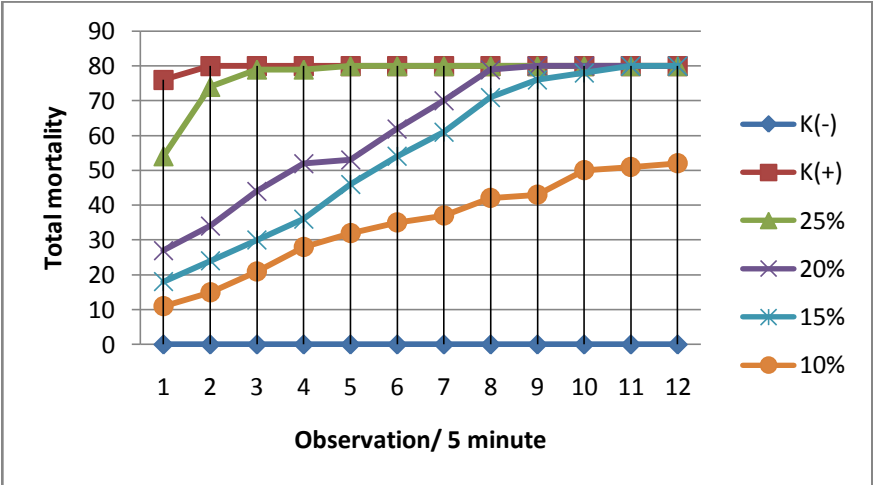
126 The *Ae. aegypti* mosquitoes died after spraying the *P. paniculata* plant extract at 10% and  
127 15% concentrations. They died slowly by flying irregularly and actively and falling in a tilted  
128 body position. The mosquitoes appeared weak, with some legs still moving, before becoming  
129 paralyzed, dying, and their bodies remaining stiff. The death was faster at 20% and 25%

130 concentrations. In the positive control (K+), death occurred in less than five minutes, while in the  
 131 negative control (K-), the *Ae. Aegypti* mosquitoes tried to avoid the spray during the four  
 132 experiment repetitions.

133  
 134 ***Ae. aegypti* Mosquitoes Death Rate at Each Concentration With Four Repetitions**

135  
 136 Based on Figure 1 In the treatment group, the *P. paniculata* plant extract was sprayed  
 137 with 10%, 15%, 20% and 25% concentration in the first 5 minutes of observation. *Ae.aegypti*  
 138 mosquitoes death rate with 4 repetitions in a row was 11, 18, 27 and 54 individuals. In the 60  
 139 minutes, the total mortality of *Ae. aegypti* mosquitoes in 4 repetitions was 52, 80, 80 and 80. In  
 140 positive control (K+) using synthetic insecticide baygon (cypermethrin) in less than 5 minutes  
 141 *Ae. aegypti* mosquitoes fell and died in 4 repetitions, namely 76 individuals. While in the  
 142 negative control (+) using distilled water did not cause death. There was 80 *Ae. aegypti*  
 143 mosquitoes were death at a concentration of 15% occurred at 51-55 minutes, a 20%  
 144 concentration of 80 total deaths occurred at 41-45 minutes and a 25% concentration of 80 total  
 145 deaths occurred at 21-25 minutes.

146



147  
 148

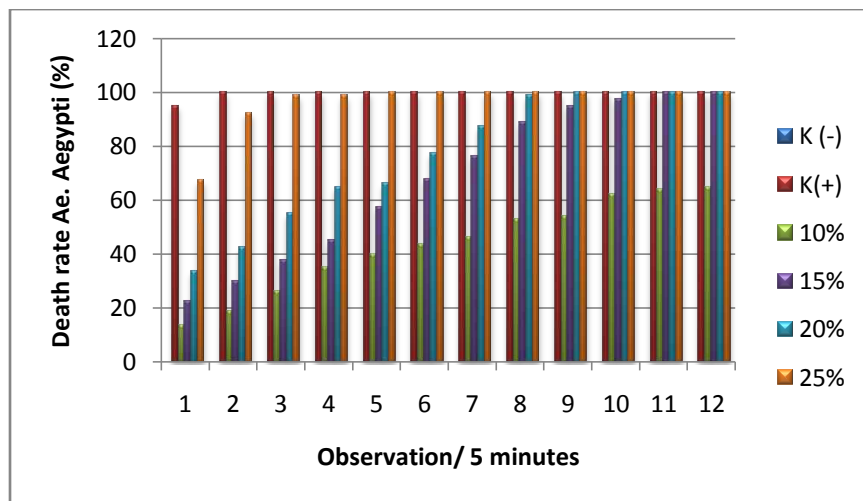
149 Figure 1 *Ae. Aegypti* mosquitoes death rate at each concentration with four repetitions  
 150 (Primary Data. 2020 )

151  
 152 **The Average of *Ae. aegypti* Death Rate at Each Concentration Every Five Minutes Observation**

153  
 154  
 155 Based on Figure 2, the treatment group was sprayed with *P. paniculata* plant extract with a  
 156 concentration of 10%, 15%, 20% and 25% in the first 5 minutes of observation, the average  
 157 mortality of *Ae. aegypti* mosquitoes with 4 repetitions in a row was 2.75 tails (13.75%), 4.5 tails  
 158 (22.5%), 6.75 tails (33.75% ) and 13 tails (67.5%). *Ae.aegypti* mosquitoes death rate in 60  
 159 minutes with 4 repetitions at a concentration of 10% was 13 tails (65%), while the concentrations  
 160 of 15%, 20% and 25% were 20 tails (100%). 95% *Ae. aegypti* mosquito fell and died in positive  
 161 control (K+) using synthetic insecticide Baygon (cypermethrin) in less than 5 minutes with 4  
 162 repetitions. While the negative control (-) using distilled water did not cause death. *Ae.Aegypti*  
 163 *mosquitos* totally died at 15% concentration in 51 until 55 minutes, 20% concentration of 80 total

164 deaths occurred at 41-45 minutes and 25% concentration of 80 total deaths occurred at 21-25  
165 minutes.

166



167

168

169 Figure 2: The Average Mortality of *Ae. aegypti* at each Concentration of Five Minutes  
170 Observation

171 Based on the results of study in figure 1 and 2, explained that the different concentrations  
172 of *P. paniculata* plant extracts gave different effects on number of *Ae. aegypti* mosquitoes death  
173 in each treatment and repetition. The number of *Ae.aegypti* mosquitoes death tends to increase  
174 along with the increase in the concentration of *P. paniculata* plant extracts. It's means that the  
175 higher used of the concentration, the higher of potency of the *P. paniculata* plant extract as a  
176 bioinsecticide against the *Ae. Aegypti* mosquito. During one hour observations showed the  
177 increasing number of *Ae. Aegypti* mosquitos's death. It's explained the longer an observation  
178 time, the greater potential as a bioinsecticide. This is supported by the results of the Kruskall-  
179 Wallis test (p-value of  $0.001 < 0.05$ ), which means that there is a significant difference between  
180 the death rate of the *Ae. aegypti* mosquito and the difference in the concentration of the *P.*  
181 *paniculata* plant extract (10%, 15%, 20%, 25 %) of Knockdown Time acceleration. The results  
182 of the Kruskall-Wallis test in this study were correct and continued with the Spearman  
183 correlation test with the results of p-value  $0.008 < 0.05$  there was a significant correlation  
184 between the increase in the concentration of the extract (treatment group) and the knockdown  
185 time of the *P. paniculata* plant extract. From the results of the Kruskall-Wallis test and the  
186 Spearman correlation test, it can be concluded that the greater of concentration, the faster the  
187 time of death of *Ae. aegypti* mosquito, the strength of the correlation is denoted by a correlation  
188 coefficient 0.312 (31.2%). Correlation coefficient occurred with positive value means that  
189 correlation is moderate. The higher concentrations of *P. paniculata* plants extract then the faster  
190 of knockdown time.

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**Insecticide Score of *P. paniculata* Plants**

Table 1 Average of *Ae. Aegypti* Falling Down At Various Treatment Concentrations With Knockdown Time<sub>50</sub> (KT<sub>50</sub>)

Time	10%	15%	20%	25%	K (+)	K(-)
5	2,75	4,5	6,75	13,5	19	0
10	3,75	6	8,5	18,5	20	0
15	5,25	7,5	11	19,75	20	0
20	7	9	13	19,75	20	0
25	8	11,5	13,25	20	20	0
30	8,75	13,5	15,5	20	20	0
35	9,25	15,25	17,5	20	20	0
40	10,5	17,75	19,75	20	20	0
45	10,75	19	20	20	20	0
50	12,5	19,5	20	20	20	0
55	12,75	20	20	20	20	0
60	13	20	20	20	20	0

Source : (Primary Data, 2020)

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Knockdown Time<sub>50</sub> (KT<sub>50</sub>) is the times to knockdown of *Ae. aegypti* mosquitoes. In Table 1, the average repetition of the study show during the 10% concentration of *P. paniculata* plant extract had a KT<sub>50</sub> between 36 until 40 minutes, 15% concentration had a KT<sub>50</sub> between 21 until 25 minutes, in the *P. paniculata* plant extract with 20% concentration the KT<sub>50</sub> was obtained between 11 and 15 minutes and in the *P. paniculata* plant extract with a concentration of 25%, the KT<sub>50</sub> was less than 5 minutes. In the average iteration of the study, there were no *Ae. aegypti* mosquitoes that fell on the negative control and the Knockdown Time<sub>50</sub> (KT<sub>50</sub>) was less than 5 minutes in the positive control. This means that the different concentrations of *P. paniculata* plant extracts gave different effects on the number of *Ae. aegypti* that fell, as well as on each treatment and repetition.

Table 2: Bio Insecticide Extract *P. paniculata* based on Knockdown Time<sub>50</sub>

Group Control	KT50 (Time)	Knockdown Effect	Insecticide Score	Interpretation
Concentration 10%	36-40	-	1	-
Concentration 15%	21-25	-	2	-
Concentration 20%	11-15	+	3	Weak Knockdown
Concentration 25%	< 5	+++	5	Quick Knockdown
Positive Control	< 5	+++	5	Quick Knockdown

Interpretation Data(+ weak Knockdown +++ Quick Knockdown)[30]  
(Primary Data, 2020)

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217 Table 2 shows the effectiveness level of the four *P. paniculata* plant extract  
218 concentrations and positive control based on Insecticide Knockdown Time<sub>50</sub> (KT<sub>50</sub>). A 10%  
219 concentration of the *P. paniculata* plant extract had a KT of<sub>50</sub> between 36-40 minutes. This  
220 implies an Insecticide Score of 1 or no knockdown effect. Similarly, a 15% concentration of the  
221 plant extracts had a KT of<sub>50</sub> between 21-25 minutes, indicating an Insecticide Score of 2 or no  
222 knockdown. A 20% concentration of the plant extract had a KT of<sub>50</sub> between 11-15 minutes,  
223 implying an Insecticide Score of 3 or a weak knockdown effect. Furthermore, a 25%  
224 concentration of *P. paniculata* plant extract had a KT<sub>50</sub> in less than 5 minutes. This indicates an  
225 Insecticide Score of 5 or a quick knockdown effect. The 25% concentration had a KT<sub>50</sub> in less  
226 than 5 minutes, similar to the positive control, with an Insecticide Score of 5 or a quick  
227 knockdown effect. Therefore, the 25% concentration of *P. paniculata* plant extract was most  
228 effective in knocking out *Ae. Aegypti* mosquitoes, with an Insecticide Score of 5 or a quick  
229 knockdown effect.

230

## 231 B. Discussion

232 The results in Figures 1 and 2 show that the number and percentage of *Ae. aegypti*  
233 mosquito deaths increased with the concentration of *P. paniculata* plant extract. Higher  
234 concentrations increase the accumulation of *P. paniculata* plant extract's toxic bioactive  
235 compounds in *A. aegypti* mosquitoes' bodies, increasing their mortality.

236 Kosini examined the effect of the *Gnidia kaussiana* (*Thymeleaceae*) extract on  
237 *Callosobruchus maculatus*. The study explained that increased absorption of toxic compounds in  
238 the *Gnidia kaussiana* extract accelerates the mortality process of larvae *Callosobruchus*  
239 *maculatus* by melanizing the cuticle. This disrupts the endocrine system due to the presence of  
240 secondary metabolites such as terpenoids, alkaloids, and flavonoids[31]. The study of da Botas et  
241 al explained that essential oil *Baccharisreticularia* DC and *limonene* as a larvacide agent on  
242 Controlling *Ae. Aegypti* (Diptera: Culicidae), it's also able to inhibit the formation of  
243 acetylcholinesterase enzyme by blocking the nerve signal of transduction which can cause the  
244 death and paralysis in *Ae. aegypti* larvae. The more larvae of *Ae. aegypti* absorbs the toxic  
245 compounds in *B. reticularia* essential oil, the more *Ae. aegypti* death. Also, the longer exposure  
246 to *B. reticularia* essential oil compounds will increase the level of toxicity.[32].

247 Many previous studies stated that the bioactive plant compounds have insecticidal,  
248 larvicidal, repellent, and environmentally-friendly effects useful for insect control. According to  
249 Suluvoy, essential oils, flavonoids, alkaloids, glycosides, esters, and fatty acids have anti-insect  
250 effects. Therefore, they could be used as an alternative to chemical compounds in insect control  
251 as repellents, feeding deterrents or antifeedants, toxicants, inhibitors, growth factors,  
252 chemosterilants, and attractants[21].

253 Previous studies explained that *Polygala* plants contain bioactive compounds with  
254 various biological activities, such as alkaloids, saponins, flavonoids, phenols, tannins, steroids,  
255 and terpenoids[27]. Tannins and flavonoids are phenolic plant compounds that act as primary  
256 antioxidants or free radical scavengers[33]. The bioactive compounds of *P. paniculata* able to  
257 affect the mortality of *Ae. aegypti* mosquitoes are flavonoids, saponins, tannins, alkaloids,  
258 steroids, and terpenoids. In this study, the *Ae. aegypti* mosquitoes death was caused by compounds  
259 in the *P. paniculata* plant extract. The compounds entered the mosquito's body through contact  
260 or respiratory poisoning and the mouth and digestive tract, causing stomach poisoning.

261 The analysis shows that the number of *Ae. aegypti* mosquitoes that died when exposed to  
262 the *P. paniculata* plant extract varied with the extract's concentration. Variations in the  
263 concentrations affected the mortality of *Ae. aegypti* mosquitoes differently in each treatment and  
264 repetition. The results in Figures 1 and 2 show that spraying a 25% concentration of the *P.*  
265 *paniculata* plant extract for 60 minutes kills 80 *Ae. Aegypti* mosquitoes in less than the first five  
266 minutes. This is the largest number of deaths that occurred faster than other concentrations.  
267 Therefore, the higher concentration of the plant extract increased its effectiveness as a vegetable  
268 insecticide against mosquitoes.

269 This study compared the effectiveness level of the four *P. paniculata* plant extract  
270 concentrations with positive controls based on Insecticide Knockdown Time<sub>50</sub> (KT<sub>50</sub>). A 25%  
271 concentration of the plant extract had a KT<sub>50</sub> of less than five minutes, implying an Insecticide  
272 Score of 5 or a quick knockdown effect. This is in line with the 2006 WHO standard, which  
273 stated that an insecticide is has a knockdown time required to drop a vector when the median  
274 knockdown ranges between 3-5[30]. Furthermore, it has a quick knockdown effect when it has a  
275 KT<sub>50</sub> of less than five minutes. According to Norris, a good insecticide requirement for  
276 controlling disease vector insect species must cause a rapid knockdown of the target species,  
277 especially active pathogens. Additionally, it must quickly intervene and kill adult mosquitoes  
278 [17].

279 The 25% concentration had a KT<sub>50</sub> of less than five minutes, similar to a positive control  
280 Baygon containing cypermethrin. They both had an Insecticide Score 5, implying a quick  
281 knockdown effect. Therefore, the 25% concentration of the *P. paniculata* plant extract was the  
282 most effective in knocking down *Ae. aegypti* mosquitoes. This is because it had an Insecticide  
283 Score of 5 or a quick knockdown effect. The positive control treatment was intended to compare  
284 the quality of *P. paniculata* plant extract concentration. In contrast, the negative control  
285 treatment was used to compare its effectiveness with the plant extract. The results showed that no  
286 mosquito died after 60 minutes of observation.

287 Chang highlighted the need to use bio-insecticides as an alternative insect controller.  
288 Using inappropriate insecticides causes insects to adapt easily by metabolic detoxification  
289 quickly and survival. This causes synchronization of insect immunity passed on to their  
290 offspring. Therefore, it is necessary to develop safe alternative insecticides, larvicides, and  
291 repellents effective for humans, animals, the environment, and the ecosystem. Natural  
292 insecticides are needed to suppress vector resistance and slow down their genetic  
293 adaptation[34]. According to Sulovoy and Grace., botanical insecticides only affect target insects,  
294 do not destroy beneficial natural enemies, and are a safe and residue-free food source. [21]. It is  
295 more environmentally friendly, effective, cheap, and naturally available[12].

## 296 **Conclusion**

297 The active substance contained in the *P. paniculata* plant extract has the ability to drop  
298 and kill *Ae. aegypti* mosquitoes. This is because more compounds in the plant extracts exposed  
299 to the mosquitoes increase the knockdown effect time. Therefore, the plant extract is a potential  
300 alternative insecticide for controlling *Ae. aegypti* mosquitoes. It does not leave residues in the  
301 environment and is safe for other living beings. Also, the extract's compounds do not cause  
302 resistance against *Ae. aegypti* mosquitoes, and the plant has economic value and is beneficial to  
303 cultivate.

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307 Mosquito" with Contract Number No. 08/STIKes-HTP/VI/2020/0147, A.

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4. Update Submitted Revised Result Minor (12 Juli 2022)

Dokumen: a. Response Letter Reviewer 1&2

b. Revised Result Minor



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Kotak Masuk 523

# MS #1303: Update submitted for "Bio-insecticide's Extract of Scented Root (Polygala paniculata) in Controlling the Mosquito Aedes aegypti (L.)"

Eksternal Kotak Masuk x



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## 4.a. Response letter Review 1 dan 2

Journal : Makara Journal of Science

Manuscript ID :

Title: **Bio-insecticides Extract of Scented Root (*Polygala paniculata*) in Controlling *Aedes aegypti* Mosquito**

There are some following issues that needs to be addressed:

Reviewer 1

<b>Reviewer's Comments</b>	<b>Author's Comments</b>
<b>Reference Number : SS21-078;</b> Comment (AB1). Please revise the title with appropriate english, vegetable insecticide is not comon to use in this title	Thank you, we have corrected the suggestion regarding the title, here are the improvements:  Bio-insecticide's Extract of Scented Root ( <i>Polygala paniculata</i> ) in Controlling <i>Aedes aegypti</i> Mosquito
Comment (AB2): It is wrong way to abbrevaiate Aedes. Please follow the rule!!!	Thank you for the suggestion regarding the correct abbreviation regarding <i>Aedes</i> to <i>Ae. aegypti</i> we have corrected everything in the manuscript regarding the title we have corrected, here are the corrections:
Comment (AB3): DHF not only problem in developing conutry. Please rewrite the paragraph	Thank you for your input regarding this paragraph, we have corrected it in the text on lines 34-37 as follows:  Indonesia is one of where dengue hemorrhagic fever (DHF) is an infectious disease and an unresolved health problem. In 2020, there were 15,132 DHF cases in Indonesia, with a death rate of 145 people and a DHF incident rate of 31.23[9].. This implies problems in efforts to control the disease.
Comment (AB4): It is not relate between vaccine, insecticide, vector. Author can delete this paragraph. Focus, please...	Thank you for your suggestion regarding there is no relationship between vaccines , insecticides, vectors we have corrected in the text on lines 38-41 as follows:  However, long-term application and extensive use of synthetic insecticides cause the accumulation of residues in food, water, soil and give adverse health effects on humans and ecosystems[10], [11]. This leaves residues that pollute the environment [3], [12]. increasing population resistance of <i>Ae. aegypti</i> [4], [5], [13]–[17].
Comment (AB5): Explain the dsitribution this plant, in the world and in Indonesia	Thank you for your suggestion to explain the distribution of this plant in the world and Indonesia, we have added it to the text on lines 57-63 as follows:

	<p><i>Polygala L.</i> is one of the largest genera belonging to the <i>Polygalaceae</i> tribe. This genus consists of 500 species and can be found in tropical, sub-tropical, temperate and mountainous areas throughout the world except New Zealand. Most of these species grow in Central and South Tropical America. Several types of <i>Polygala L.</i> that can be used as medicine such as :<i>Polygala chinensis L.</i>, <i>Polygala paniculata L.</i>, <i>Polygala polifoliaPresl.</i>, and <i>Polygala sibirica L.</i>[24].<i>Polygala paniculata</i>, also known as vetiver in Riau, Indonesia, is a good-smelling annual herbaceous plant belonging to the family Polygalaceae and the genus Polygala.</p>
<p>Comment (AB6): Use good english please</p>	<p>Thank you for We have corrected the correction regarding the subtitles to use good English in the text on line 91 as follows</p> <p>Extraction Process</p>
<p>Comment (AB7): Explain the kind of testing</p>	<p>Thank you for the correction regarding the type of test we have corrected and added to the text on line 100-109 as follows:</p> <p><b>Bio-insecticide’s Extract of <i>P. paniculata</i> Test against <i>Ae. Aegypti</i> Mosquitos</b></p> <p>Bio-insecticide’s extract of <i>P. paniculata</i> tests against <i>Ae. aegypti</i> was carried out by transferring 20 <i>Ae. aegypti</i> in each test boxes, then sprayed with various concentrationsof <i>P. paniculata</i> plant extract with four repetitions, as well as for K (+) and K(-).The effect of the concentrations on the <i>A. aegypti</i> mosquito was observed by looking at the changes in behavior, movement, and physical condition until death. <i>Ae. Aegypti</i> mosquitoes death were counted every five minutes for one hour. Moreover, the insecticide score of <i>P. paniculata</i> was determined from the number of <i>Ae. Aegypti</i> mosquitoes considered deadat 5-minute intervals. <i>Ae. Aegypti</i> mosquitoes that remained alive were left to die or killed with Baygon.</p>
<p>Comment (AB9): Use good English please</p>	<p>Thank you for the correction to use correct English, we have corrected the text on line 134-143 as follows:</p> <p>Based on Figure 1 In the treatment group, the <i>P. paniculata</i> plant extract was sprayed with 10%, 15%, 20% and 25% concentration in the first 5 minutes of observation. <i>Ae.aegypti</i> mosquitoes death rate with 4 repetitions in a row was 11, 18, 27 and 54 individuals. In the 60 minutes, the total mortality of <i>Ae. aegypti</i> mosquitoes in 4 repetitions was 52, 80, 80 and 80. In positive control (K+) using synthetic insecticide baygon (cypermethrin) in less than 5 minutes <i>Ae. aegypti</i> mosquitoes fell and died in 4 repetitions, namely 76 individuals.While in the negative control (+) using distilled water did not cause death.There was 80 <i>Ae. aegypti</i></p>

mosquitoes were death at a concentration of 15% occurred at 51-55 minutes, a 20% concentration of 80 total deaths occurred at 41-45 minutes and a 25% concentration of 80 total deaths occurred at 21-25 minutes.

Comment (AB10): Not clear, please make in 2 dimension graph

Thank you for the correction regarding the three (3) dimensional graph we have corrected to a two (2) dimensional graph in the text on lines 162-168 as follows:

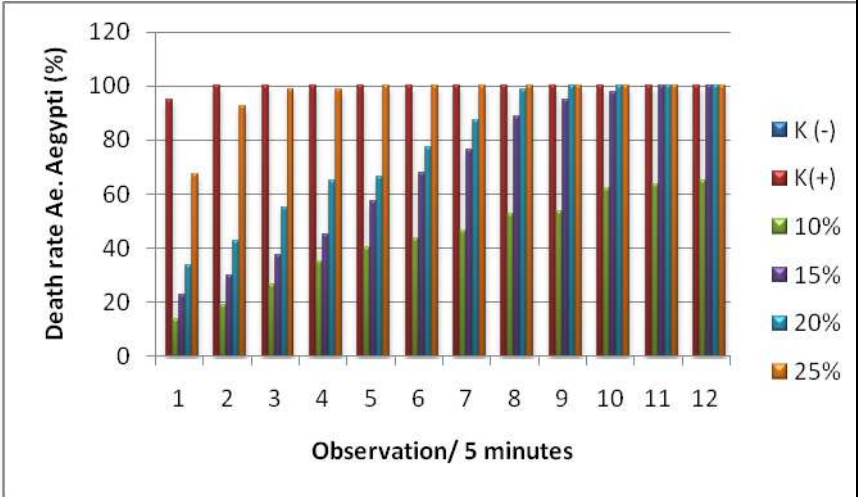


Figure 2: The Average Mortality of *Ae. aegypti* at each Concentration of Five Minutes Observation

Comment (AB11); (AB12); (AB13): No need to put the statistical analysis in here. Write as result please

Thank you for the correction to not include statistical analysis, we have changed it to include only the results in the text on lines 169-188 as follows:

Based on the results of study in figure 1 and 2, explained that the different concentrations of *P. paniculata* plant extracts gave different effects on number of *Ae. aegypti* mosquitoes death in each treatment and repetition. The number of *Ae.aegypti* mosquitoes death tends to increase along with the increase in the concentration of *P. paniculata* plant extracts. It's means that the higher used of the concentration, the higher of potency of the *P. paniculata* plant extract as a bioinsecticide against the *Ae. Aegypti* mosquito. During one hour observations showed the increasing number of *Ae. Aegypti* mosquitos's death. It's explained the longer an observation time, the greater potential as a bioinsecticide. This is supported by the results of the Kruskal-Wallis test (p-value of 0.001 <0.05), which means that there is a significant difference between the death rate of the *Ae. aegypti* mosquito and the difference in the concentration of the *P. paniculata* plant extract (10%, 15%, 20%, 25 %) of Knockdown Time acceleration. The results of the Kruskal-Wallis test in this study were correct and continued with the Spearman correlation test with the results of p-value 0.008 <0.05 there was a significant correlation between the increase in the concentration



of the extract (treatment group) and the knockdown time of the *P. paniculata* plant extract. From the results of the Kruskal-Wallis test and the Spearman correlation test, it can be concluded that the greater of concentration, the faster the time of death of *Ae. aegypti* mosquito, the strength of the correlation is denoted by a correlation coefficient 0.312 (31.2%). Correlation coefficient occurred with positive value means that correlation is moderate. The higher concentrations of *P. paniculata* plants extract then the faster of knockdown time

Comment (AB14): Please make the table more clear. Explain the highlighting data!

Thanks for the suggestion we have corrected the explanation on lines 194-198 in the manuscript

Table 1 Average of *Ae. Aegypti* Falling Down At Various Treatment Concentrations With Knockdown Time<sub>50</sub> (KT<sub>50</sub>)

Time	10%	15%	20%	25%	K (+)	K (-)
5	2,75	4,5	6,75	13,5	19	0
10	3,75	6	8,5	18,5	20	0
15	5,25	7,5	11	19,75	20	0
20	7	9	13	19,75	20	0
25	8	11,5	13,25	20	20	0
30	8,75	13,5	15,5	20	20	0
35	9,25	15,25	17,5	20	20	0
40	10,5	17,75	19,75	20	20	0
45	10,75	19	20	20	20	0
50	12,5	19,5	20	20	20	0
55	12,75	20	20	20	20	0
60	13	20	20	20	20	0

: *Ae. Aegypti* Knockdown Time(KT<sub>50</sub>);

Comment (AB15) untuk tabel 5. What mean +?

Thank you we tried to explain Regarding the meaning of +  
In table 5 : Insecticide Score of *P. paniculata* Plant Extract Based on KT<sub>50</sub>.

The knockdown effect has been defined by WHO 2006 in the book Guidelines for Testing Mosquitos Adulticides for Indoor Residual Spraying and Treatment of Mosquitos Nets  
That: Insecticide Score Based on KT 50

KT50 (minute)	Score	Knockdown effect	Interpretation
>50	0	-	-
31-49	1	-	-
16-30	2	-	-
11-15	3	+	Weak Knockdown
5-10	4	++	Strong Knockdown
<5	5	+++	Quick Knockdown

source: WHO, 2006

A value of 3 means the median knockdown is in the range of 11-15 minutes which is interpreted to have a knockdown

	<p>effect (+) but is weak. A value of 4 means the median knockdown is in the range of 5-10 minutes which is interpreted to have a strong knockdown effect (++). A value of 5 means the median knockdown is in the range of less than 5 minutes which is interpreted to have a knockdown effect (+++) that the insecticide has a “Quick Knockdown Effect” (WHO, 2006).</p> <p>So the + sign means explaining the Knockdown effect status of Bioinsecticide</p>
<p>Comment (AB16): Please change the scopus non-indexed reference to the scopus indexed reference</p>	<p>Thank you for the correction to change all non-Scopus references to Scopus-indexed references, all of which we have changed to Scopus-indexed references in the script as follows</p> <p><b>References</b></p> <p>[1] L. H. Guimarães de Oliveira <i>et al.</i>, “Agave sisalana extract induces cell death in <i>Aedes aegypti</i> hemocytes increasing nitric oxide production,” <i>Asian Pac. J. Trop. Biomed.</i>, vol. 6, no. 5, pp. 396–399, May 2016.</p> <p>[2] D. A. Fernandes <i>et al.</i>, “Larvicidal compounds extracted from <i>helicteres velutina</i> K. Schum (Sterculiaceae) evaluated against <i>aedes aegypti</i> L.,” <i>Molecules</i>, vol. 24, no. 12, p. 2315, 2019.</p> <p>[3] L. Scalvenzi <i>et al.</i>, “Larvicidal activity of <i>Ocimum campechianum</i>, <i>Ocotea quixos</i> and <i>Piper aduncum</i> essential oils against <i>Aedes aegypti</i>,” <i>Parasite</i>, vol. 26, no. 23, pp. 1–8, 2019.</p> <p>[4] D. Valle, D. F. Bellinato, P. F. Viana-Medeiros, J. B. P. Lima, and A. D. J. Martins Junior, “Resistance to temephos and deltamethrin in <i>aedes aegypti</i> from Brazil between 1985 and 2017,” <i>Mem. Inst. Oswaldo Cruz</i>, vol. 114, no. 3, pp. 1–17, Mar. 2019.</p> <p>[5] Z. H. Amelia-yap, C. D. Chen, M. Sofian-azirun, and V. L. Low, “Pyrethroid resistance in the dengue vector <i>Aedes aegypti</i> in Southeast Asia : present situation and prospects for management,” <i>Parasit. Vectors</i>, vol. 11, no. 332, pp. 1–17, 2018.</p> <p>[6] S. Leta, T. Jibat, E. M. De Clercq, K. Amenu, M. U. G. Kraemer, and C. W. Revie, “International Journal of Infectious Diseases Global risk mapping for major diseases transmitted by <i>Aedes aegypti</i> and <i>Aedes albopictus</i>,” <i>Int. J. Infect. Dis.</i>, vol. 67, pp. 25–35, 2018.</p> <p>[7] M. Y. Lee, “Essential Oils as Repellents against</p>

	<p>Arthropods,” <i>Biomed Res. Int.</i>, vol. -, no. -, pp. 1–9, 2018.</p> <p>[8] WHO, “Dengue and severe dengue,” <i>World Health Organization</i>, 19-May-2021. [Online]. Available: <a href="https://www.who.int/news-room/fact-sheets/detail/dengue-and-severe-dengue">https://www.who.int/news-room/fact-sheets/detail/dengue-and-severe-dengue</a>. [Accessed: 06-Dec-2021].</p> <p>[9] Kemenkes RI, <i>Incidence Rate Per 100.000 Penduduk Demam Berdarah Dengue Tahun 2010-2018</i>. Jakarta, 2020.</p> <p>[10] K. Baana, H. Angwech, and G. M. Malinga, “Ethnobotanical survey of plants used as repellents against housefly, <i>Musca domestica</i> L. (Diptera: Muscidae) in Budondo Subcounty, Jinja District, Uganda,” <i>J. Ethnobiol. Ethnomed.</i>, vol. 14, no. 1, pp. 1–8, May 2018.</p> <p>[11] A. T. H. Mossa, S. M. M. Mohafrash, and N. Chandrasekaran, “Safety of natural insecticides: Toxic effects on experimental animals,” <i>Biomed Res. Int.</i>, vol. 2018, pp. 1–18, 2018.</p> <p>[12] E. A. Rohmah, S. Subekti, and M. Rudyanto, “Larvicidal Activity and Histopathological Effect of <i>Averrhoa bilimbi</i> Fruit Extract on <i>Aedes aegypti</i> from Surabaya, Indonesia,” <i>J. Parasitol. Res.</i>, vol. 2020, pp. 1–5, Aug. 2020.</p> <p>[13] C. S. Bibbs, D. A. Hahn, P. E. Kaufman, and R. Xue, “Sublethal effects of a vapour-active pyrethroid, transfluthrin, on <i>Aedes aegypti</i> and <i>Ae. albopictus</i> (Diptera: Culicidae) fecundity and oviposition behaviour,” <i>Parasit. Vectors</i>, vol. 11, no. 1, p. 486, Dec. 2018.</p> <p>[14] P. A. Rojas-Pinzón, . Juan, J. Silva-Fernández, and J. Dussán, “Laboratory and simulated-field bioassays for assessing mixed cultures of <i>Lysinibacillus sphaericus</i> against <i>Aedes aegypti</i> (Diptera: Culicidae) larvae resistant to temephos,” <i>Applied Entomol. Zool.</i>, vol. 53, pp. 183–191, 2018.</p> <p>[15] E. L. R. De Sá <i>et al.</i>, “Evaluation of insecticide resistance in <i>aedes aegypti</i> populations connected by roads and rivers: The case of tocantins state in Brazil,” <i>Mem. Inst. Oswaldo Cruz</i>, vol. 114, no. 2, pp. 1–10, Feb. 2019.</p> <p>[16] S. Murfadunnisa <i>et al.</i>, “Larvicidal and enzyme inhibition of essential oil from <i>Spheranthus amaranthoides</i> (Burm.) against lepidopteran pest <i>Spodoptera litura</i> (Fab.) and their impact on non-target earthworms,” <i>Biocatal. Agric.</i></p>
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	<p><i>Biotechnol.</i>, vol. 21, no. 101324, pp. 1–9, 2019.</p> <p>[17] E. J. Norris, A. D. Gross, L. C. Bartholomay, and J.R. Coats, “Plant essential oils synergize various pyrethroid insecticides and antagonize malathion in <i>Aedes aegypti</i>,” <i>Med. Vet. Entomol.</i>, vol. 33, pp. 453–466, 2019.</p> <p>[18] J. G. Galvão <i>et al.</i>, “<i>Lippia gracilis</i> essential oil in <math>\beta</math>-cyclodextrin inclusion complexes: an environmentally safe formulation to control <i>Aedes aegypti</i> larvae,” <i>Pest Manag. Sci.</i>, vol. 75, no. 2, pp. 452–459, Feb. 2019.</p> <p>[19] Y. Kandel <i>et al.</i>, “Widespread insecticide resistance in <i>Aedes aegypti</i> L. From New Mexico, U.S.A.,” <i>PLoS One</i>, vol. 14, no. 2, pp. 1–16, Feb. 2019.</p> <p>[20] G. N. Pandiyan, N. Mathew, and S. Munusamy, “Larvicidal activity of selected essential oil in synergized combinations against <i>Aedes aegypti</i>,” <i>Ecotoxicol. Environ. Saf.</i>, vol. 174, pp. 549–556, 2019.</p> <p>[21] J. K. Suluvoy and V. M. Berlin Grace, “Phytochemical profile and free radical nitric oxide (NO) scavenging activity of <i>Averrhoa bilimbi</i> L. frui extract,” <i>3 Biotech</i>, vol. 7, no. 1, pp. 1–11, May 2017.</p> <p>[22] H. D. Manh, D. T. Hue, N. Thi, T. Hieu, D. Thi, and T. Tuyen, “The Mosquito Larvicidal Activity of Essential Oils from <i>Cymbopogon</i> and <i>Eucalyptus</i> Species in Vietnam,” <i>MPDI</i>, vol. 11, no. 128, pp. 1–7, 2020.</p> <p>[23] B. Sisay, T. Tefera, M. Wakgari, G. Ayalew, and E. Mendesil, “The efficacy of selected synthetic insecticides and botanicals against fall armyworm, <i>spodoptera frugiperda</i>, in maize,” <i>Insects</i>, vol. 10, no. 2, pp. 1–14, Feb. 2019.</p> <p>[24] J. L. C. . Van Valkenburg, <i>Plant Resources of south-east asia 12 (2) Medicinal and poisonous plant 2</i>. Bogor, Indonesia: Bogor Prosea, 2002.</p> <p>[25] F. da R. Lapa <i>et al.</i>, “Antinociceptive Properties of the Hydroalcoholic Extract and the Flavonoid Rutin Obtained from <i>Polygala paniculata</i> L. in Mice,” <i>Basic Clin. Pharmacol. Toxicol.</i>, vol. 104, no. 4, pp. 306–315, Apr. 2009.</p> <p>[26] M. Farina <i>et al.</i>, “Protective effects of <i>Polygala paniculata</i> extract against methylmercury- induced neurotoxicity in</p>
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	<p>mice,” <i>J. Pharm. Pharmacol.</i>, pp. 1503–1508, 2005.</p> <p>[27] J. L. C. Da Rocha, R. F. A. De Tripodi Calumby, D. F. Da Silva, H. N. Brandão, C. F. Villarreal, and F. O. De Lima, “Evaluation of biological activity of polygala boliviensis in experimental models,” <i>Iran. J. Pharm. Res.</i>, vol. 18, no. 2, pp. 793–802, Mar. 2019.</p> <p>[28] J. D. Fontana <i>et al.</i>, “Accelerating the Morphogenetic Cycle of the Viral Vector <i>Aedes aegypti</i> Larvae for Faster Larvicidal Bioassays,” <i>Biomed Res. Int.</i>, vol. 2020, 2020.</p> <p>[29] Kemenkes RI, “Farmakope Herbal Indonesia,” II., Jakarta, 2017.</p> <p>[30] WHO, <i>Mosquito Adulticides for Indoor Residual Spraying and Treatment of Mosquito Nets</i>. Geneva: World Health Organization, 2006.</p> <p>[31] D. Kosini and E. N. Nukenine, “Bioactivity of novel botanical insecticide from gnidia kaussiana (Thymeleaceae) against callosobruchus maculatus (Coleoptera: Chrysomelidae) in stored vigna subterranea (Fabaceae) grains,” <i>J. Insect Sci.</i>, vol. 17, no. 1, pp. 1–7, 2017.</p> <p>[32] G. S. da Botas <i>et al.</i>, “molecules Baccharis reticularia DC. and Limonene Nanoemulsions: Promising Larvicidal Agents for <i>Aedes aegypti</i> (Diptera: Culicidae) Control,” <i>mdpi.com</i>, vol. 22, no. -, pp. 2–14, 2017.</p> <p>[33] I. Sina, Zaharah, and M. S. M. Sabri, “Larvicidal activities of extract flower averrhoa bilimbi L. Towards important species mosquito, anopheles barbirostris (diptera: Culicidae),” <i>Int. J. Zool. Res.</i>, vol. 12, no. 1–2, pp. 25–31, 2016.</p> <p>[34] X. Chang <i>et al.</i>, “Multiple Resistances and Complex Mechanisms of <i>Anopheles sinensis</i> Mosquito: A Major Obstacle to Mosquito-Borne Diseases Control and Elimination in China,” <i>PLoS Negl. Trop. Dis.</i>, vol. 8, no. 5, p. e2889, May 2014.</p>

Reviewer 2

<b>Reviewer's Comments</b>	<b>Author's Comments</b>
<p>1. Consider using <i>Ae. aegypti</i> as an abbreviation for <i>Aedes aegypti</i>.</p>	<p>Thank you for providing corrections to change the abbreviation <i>Aedes aegypti</i> to <i>Ae. Aegypti</i>, we have corrected this suggestion in the manuscript</p>
<p>2. The abbreviations YFV, ZIKV, CHIKV and DENV are used to refer to viruses (etiological agents) and not to diseases. Consider tweaking your text (page 01)</p>	<p>Thank you for the suggestion to consider changing the text regarding the abbreviations YHF, ZIKV, CHIKV and DENV in paragraph 1. We have changed this in the text on lines 23-25 as follows:</p> <p>The <i>Aedes aegypti</i> mosquito (L) belongs to the Order Diptera and family Culicidae, the main vector often neglected as a transmitter of diseases in humans. The diseases include Yellow Fever, Zica, Chikungunya, Dengue Haemorrhagic Fever and other arboviruses[1]–[4].</p>
<p>3. There is redundancy between the Materials and Methods topic presented in the subtopics "Preparation of Materials and Tools" and "Extract Making". The text needs to be adjusted to avoid such redundancies (page 2-3).</p>	<p>Thank you for the correction</p> <p>Regarding the redundancy between the topic of Materials and Methods presented in the subtopic Preparation of Materials and Tools and Preparation of Extracts in the text, we have corrected it on lines 78-83 in the text as follows:</p> <p><b>Preparation of Materials and Tools</b></p> <p>This study used 2500 gram <i>P. paniculata</i>, 5 litre of 96% ethanol, 5 litre of distilled water, 1 bottle of synthetic insecticide Baygon (cypermethrin), and 480 <i>Ae. aegypti</i> mosquito. The equipment used in this study includes a blender, analytical scales, Rotary Vacuum Evaporator, stopwatch, shaker water bath, thermometer, hygrometer, Buchner funnel, stick, basin, test box, syringe, and a spray bottle.</p>
<p>4. In Figure 1, only one caption should be kept for an image. The descriptions that are in the figure must be placed in the legend (page 4).</p>	<p>Thank you for the correction regarding the information in Figure 1, we have corrected it on line 144-147 as follows:</p>

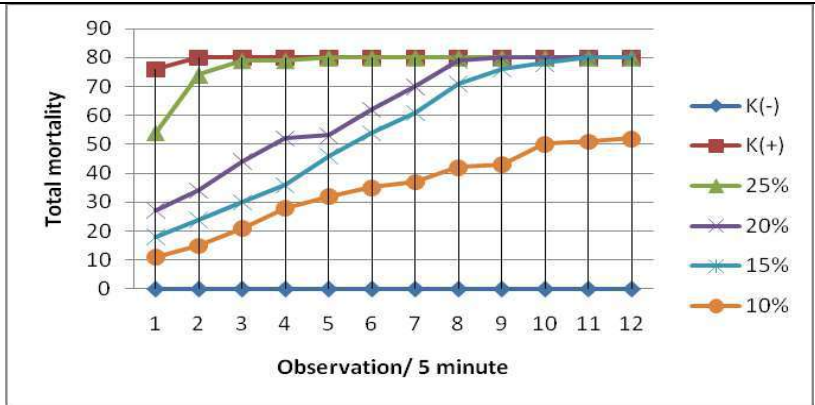


Figure 1 *Ae. Aegypti* mosquitoes total mortality at each concentration with four repetitions

5. When presenting the data in Figure 01, it is interesting to consider the average number of deaths by concentration and not the sum of deaths (Line 135).

Thank you for the advice. Figure 01 is retained but the average mortality is made in Figure 02 as follows: on lines 144-147 and lines 164-168

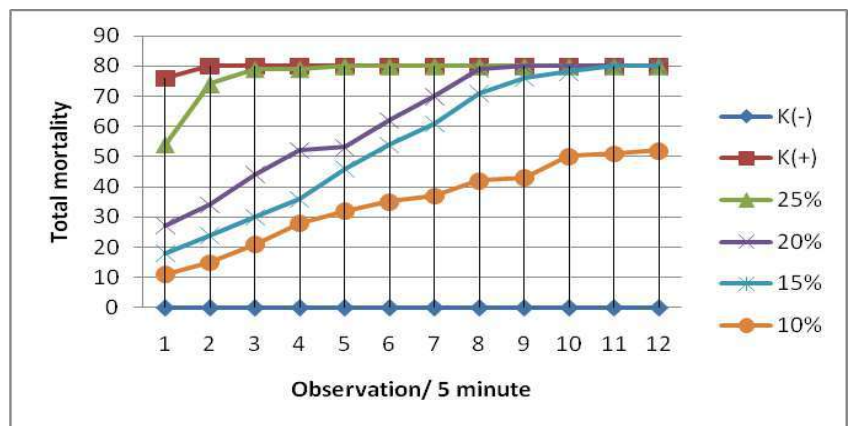


Figure 1 *Ae. Aegypti* mosquitoes total mortality at each concentration with four repetitions

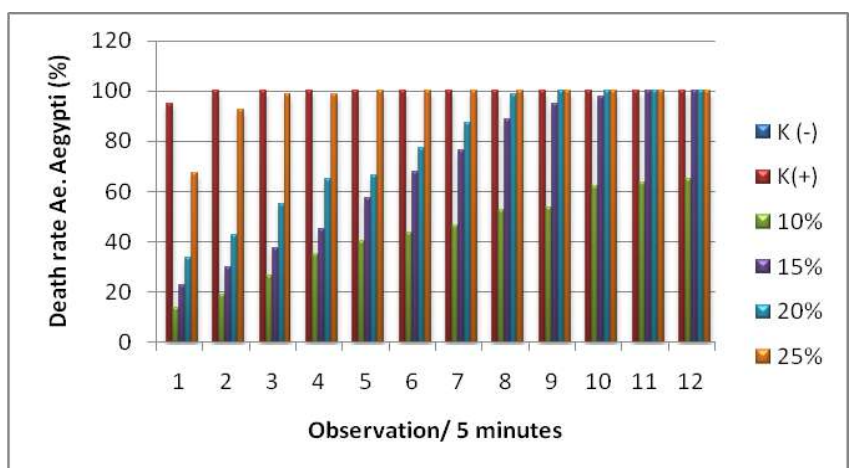


Figure 2: The Average Mortality of *Ae. aegypti* at each Concentration of Five Minutes Observation

6. Note the writing of

Thank you for the advice regarding scientific writing *Aedes*

<p>the scientific name "<i>Aedes aegypti</i>" on pages 5 and 6.</p>	<p>aegypti " on pages 5 and 6 we have corrected it as suggested</p>
<p>7. In table 01 the date presented in the table (0.000) are different from those discussed in the text (0.001) (line 167)</p>	<p>Thank you for the advice. Based on input from other reviewers that this table is not needed but the results from the table. This is conveyed in the script narrative only on lines 169-188 as follows:</p> <p>Based on the results of study in figure 1 and 2, explained that the different concentrations of <i>P. paniculata</i> plant extracts gave different effects on number of <i>Ae. aegypti</i> mosquitoes death in each treatment and repetition. The number of <i>Ae.aegypti</i> mosquitoes death tends to increase along with the increase in the concentration of <i>P. paniculata</i> plant extracts. It's means that the higher used of the concentration, the higher of potency of the <i>P. paniculata</i> plant extract as a bioinsecticide against the <i>Ae. Aegypti</i> mosquito. During one hour observations showed the increasing number of <i>Ae. Aegypti</i> mosquitos's death. It's explained the longer an observation time, the greater potential as a bioinsecticide. This is supported by the results of the Kruskall-Wallis test (p-value of 0.001 &lt;0.05), which means that there is a significant difference between the death rate of the <i>Ae. aegypti</i> mosquito and the difference in the concentration of the <i>P. paniculata</i> plant extract (10%, 15%, 20%, 25 %) of Knockdown Time acceleration. The results of the Kruskall-Wallis test in this study were correct and continued with the Spearman correlation test with the results of p-value 0.008 &lt;0.05 there was a significant correlation between the increase in the concentration of the extract (treatment group) and the knockdown time of the <i>P. paniculata</i> plant extract. From the results of the Kruskall-Wallis test and the Spearman correlation test, it can be concluded that the greater of concentration, the faster the time of death of <i>Ae. aegypti</i> mosquito, the strength of the correlation is denoted by a correlation coefficient 0.312 (31.2%). Correlation coefficient occurred with positive value means that correlation is moderate. The higher concentrations of <i>P. paniculata</i> plants extract then the faster of knockdown time</p>
<p>8. The value 0.08 is greater than 0.05, not less. (line 185)</p>	<p>Thanks for the correction. We have deleted this information because the related table has been changed to a new narrative</p>
<p>9. The Time required for 50% knockdown of mosquitoes (KT50) could be accurately estimated using probit analysis (table 4).</p>	<p>Thank you for the suggestion, to use probit analysis to determine the time required for 50% mosquito knockdown (KT50) in Table 4  We try to explain  Knockdown time (KT50) means the time it takes for each concentration of <i>P. paniculata</i> Bioinsecticide to drop 50% of the average <i>Ae. aegypti</i>. We did not use the probit analysis in Table 4, because here we only looked at how many minutes the average number of mosquitoes fell by 50% for each concentration of <i>P. paniculata</i> Bioinsecticide.</p>



<p>10. In obtaining table 05, were the mortality averages considered?</p>	<p>Thank you for the correction.</p> <p>We try to explain, We have changed Table 05 to Table 02 because Tables 01, 02, 03 have been omitted according to input from other reviewers. For Table 02, we do not use the average mortality, but we use <math>KT_{50}</math> (Time) which is the time it takes to drop 50% of <i>Ae. aegypti</i> after being sprayed with <i>P. paniculata</i> Bioinsecticide at each test concentration.</p>
<p>11. In line 196 refers to table 04 and in line 207 refers to table 05.</p>	<p>Thank you for the correction. We have corrected the script on line 201-202 as follows:</p> <p>Knockdown Time<sub>50</sub> (<math>KT_{50}</math>) is the times to knockdown of <i>Ae. aegypti</i> mosquitoes. In Table 1, the average repetition of the study show during the 10% concentration of <i>P. paniculata</i> plant</p>
<p>12. According to the presentation of the text, I suggest that the Results section be presented separately from the Discussion section.</p>	<p>Thank you for the advice. We have corrected the manuscript according to the following suggestions:</p> <p>A. Results pada line 117 B. Discussion pada line 229</p>

Reviewer 3

Reviewer's Comments	Author's Comments

5. Proofreading (4 Juli 2022)

Dokumen: a. Tracked proofreading

b. Respon letter-Proofreading

## 5.a. Tracked Proofreading

### Bio-insecticide's Extract of Scented Root (*Polygala paniculata*) in Controlling The Mosquito *Aedes aegypti* Mosquito (L.)

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#### ABSTRACT

Controlling *Aedes aegypti* mosquitoes with chemical insecticides causes resistance ~~effects~~ on humans, environmental residues, ~~as well as and~~ contaminates food and water. *Polygala paniculata* ~~has is a~~ potential ~~as an~~ alternative ~~of insecticide to insecticides~~ in controlling *Ae. Aegypti* mosquitoes. This study ~~further~~ aimed to determine the effect of the Insecticide Score of *P. paniculata* ~~extract extracts~~ on *Ae. Aegypti* mosquitoes' ~~the~~ mortality of *Ae. aegypti* mosquitoes based on  $KT_{50}$ . ~~The A total of~~ 20 mosquitoes ~~for each concentration~~ were used to examine the effectiveness of 10%, 15%, 20%, and 25% ~~concentration~~ of *P. paniculata* ~~concentration with extracts compared to a~~ positive and a negative ~~controls in control with~~ four repetitions every five minutes for one hour replications. The results showed ~~the effect of extract~~ that *P. paniculata* ~~extracts had an effect on Aeaegypti mosquitoes mosquito~~ mortality. ~~The There~~ were significant differences in mortality rate between concentrations (Kruskal-Wallis test resulted in a  $p$ -value of  $\leq 0.001 < 0.05$ , while the Spearman Correlation test gave a  $p$ -value of  $0.008 < 0.05$ . ~~Furthermore, the~~ Similarly, there was a weak but significant correlation ~~strength~~ ~~was +between plant extract concentration and mortality (Spearman correlation:  $r = +0.312$ ,  $p = 0.312$  or 31.2%, with 0.008).~~ The 10% ~~concentration, while and~~ 15% concentrations had  $KT_{50}$  with Insecticide Scores of 1 and 2, ~~respectively~~, implying ~~that both had~~ no knockdown effect. A 20% concentration had  ~~$KT_{50}$  with a  $KT_{50}$  with~~ an Insecticide Score of 3, indicating a weak knockdown effect, ~~while whereas~~ a 25% concentration had  $KT_{50}$  with an Insecticide Score of 5, signifying a quick knockdown ~~time in A time on Ae. Aegypti~~ mosquitoes. Therefore, extract *P. paniculata* ~~may have the extract at a concentration of 25% has a potential~~ ~~for use~~ as a bio-insecticide ~~in insecticide in~~ controlling *Ae. aegypti* mosquitoes.

Keywords: extract *P. paniculata*, *Aedes aegypti* mosquito, bio-insecticides

#### 1. Introduction

The *Aedes aegypti* mosquito (L.), which belongs to the Order Diptera and family Culicidae, is the main vector, often neglected ~~as a transmitter of, transmitting human~~ diseases ~~in humans~~. ~~The diseases include including~~ Yellow Fever, Zica, Chikungunya, -Dengue Haemorrhagic Fever and other arbo viruses [1-4]. ~~The, Aedes aegypti transmits the~~ human arboviral disease

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Comment [Editor1]: Remark: It is standard practice to indicate the authority the first time you mention a species.

Comment [Editor2]: Remark: Please check your target journal guidelines for the correct style of citing references within text. Put a space between the last word and the open bracket. Usually, references are cited like this: [1-4], not like this: [1] - [4].

43 | ~~transmitted by *Ae. aegypti* is~~, a global public health threat[5]. ~~This disease that~~ causes significant  
44 | morbidity and mortality in developing countries [1]. The transmission of dengue fever is  
45 | increasing in urban and semi-urban areas in tropical countries worldwide [6]. It is estimated that  
46 | 40% or 50–~~to~~ 528 million people worldwide are at risk of becoming infected with dengue fever  
47 | and around 10,000–20,000 people die yearly [7]. According to WHO, about 390 million cases of  
48 | dengue virus infection occur every year, of which 96 million manifest clinically with high  
49 | severity. Furthermore, WHO reports that about 3.9 billion people are at risk of being infected  
50 | with the dengue virus [8].

51 | Indonesia is one of ~~the countries~~ where dengue hemorrhagic fever (DHF) ~~occurs and~~ is an  
52 | ~~infectious disease and an~~ unresolved health problem. In 2020, there were 15,132 DHF cases ~~and~~  
53 | ~~145 deaths~~ in Indonesia, ~~with a death rate of 145 people~~ and a DHF ~~incident rate of 31.23 %~~ [9].  
54 | ~~This implies; indicating~~ problems in ~~efforts to control~~ ~~controlling~~ the disease.

55 | ~~However, long~~ Long-term application and extensive use of synthetic insecticides cause ~~the~~  
56 | accumulation of residues in food, water, ~~and~~ soil, ~~and~~ ~~give adverse~~ ~~affect human and ecosystem~~  
57 | health ~~effects on humans and ecosystems~~ [10], [11]. ~~This~~ Use of insecticides leaves residues that  
58 | pollute the environment [3], [12]. ~~increasing population and cause~~ resistance ~~of in~~ *Ae.*  
59 | ~~aegypti~~ *Aegypti* populations [4], [5], [13]–[17].

60 | Controlling *Ae. aegypti* mosquitoes using synthetic insecticides involves fumigation with  
61 | pyrethroids and larvacides containing temephos [18]. ~~Pyrethroid *Aedes aegypti* develops~~  
62 | resistance ~~causes *Ae. aegypti* to keep growing, pyrethroid and multiply rapidly~~ increasing the  
63 | incidence of dengue fever (DF) worldwide. This ~~increases the risk of puts~~ almost half of the  
64 | world's population ~~at the risk of~~ being infected with the disease [5]. ~~Similarly, using~~ Using  
65 | temephos to kill larvae ~~increases the resistance of mosquitoes larvae, *A. aegypti* has a similar~~  
66 | ~~effect~~. This has been reported in several countries, such as Brazil [4], Mexico [19] and ~~Southeast~~  
67 | ~~Asia~~—countries ~~in south-east Asia~~, including Indonesia, Malaysia, Philippines, Thailand,  
68 | Singapore, Laos, and Myanmar [5].

69 | There is a need to obtain alternative insecticides effective in controlling the ~~mosquito~~ *A.*  
70 | ~~aegypti~~ population *A. aegypti* [20]. This could be achieved ~~by~~ using natural plant chemical  
71 | compounds ~~with the potential to control the mosquito population effectively. The that are~~  
72 | environmentally friendly ~~method would guarantee plant protection~~ and avoid the side effects of  
73 | synthetic insecticides; [10], [21], [22]. Bioactive plant compounds are biodegradable,  
74 | environmentally friendly, and non-toxic to ~~other non-target~~ insects [2]. ~~This is because bio-~~  
75 | ~~insecticides~~ They only affect the target insect without destroying beneficial natural enemies.  
76 | ~~Additionally~~ Generally, they are a safe, economical, target-specific, biodegradable, and residue-  
77 | free ~~environmental food source~~ [23].

78 | *Polygala L.* is one of the largest genera belonging to the Polygalaceae tribe. This genus  
79 | consists of 500 species and can be found in tropical, sub-tropical, temperate, and mountainous  
80 | areas throughout the world except New Zealand. Most of these species grow in Central and  
81 | South Tropical America. ~~Several types~~ There are several species of *Polygala L.* that can be used  
82 | as medicine ~~such as including~~ *Polygala chinensis L.*, *Polygala paniculata L.*, *Polygala*  
83 | ~~polifolia~~ *Prestipolifolia Presl.*, and *Polygala sibirica L.* [24]. *Polygala paniculata*, also known as  
84 | vetiver in Riau, Indonesia, is a good-smelling annual herbaceous plant belonging to the family  
85 | Polygalaceae ~~and the genus *Polygala*~~. The plant is often used ~~as in~~ traditional medicine, ~~as~~  
86 | a tonic, and ~~in inflammation for controlling inflammatory~~ cases of asthma, bronchitis, arthritis, and  
87 | ~~other pathologies, as well as~~ kidney disorders [25]. Moreover, *P. paniculata* is used for ~~in-~~ *in-vivo*  
88 | protection against the neurotoxic effects of ~~Methylmercury~~ *methylmercury* (Hg<sup>2+</sup>) [26], bronchitis,

**Comment [Editor3]:** Remark: Please check this this. I think incidence rate should be reported as a percentage or decimal.

**Comment [Editor4]:** Remark: It is obvious from the species name that it belongs to this genus. Hence, we have deleted it.

89 | neurahenia, inflammation, amnesia, topical anesthetic, and expectorant drugs [27]. ~~The~~ *Polygala*  
90 | ~~extract produce extracts contain~~ secondary metabolites, including alkaloids, saponins,  
91 | flavonoids, phenols, tannins, steroids, and terpenoids [27]. ~~Therefore, these~~ These bioactive  
92 | compounds make *P. paniculata* potentially useful as a bio-insecticides. ~~This becomes an~~  
93 | ~~interesting study topic because the~~ insecticide. The bioactive compounds extracted from *P.*  
94 | *paniculata* have ~~never not yet~~ been explored as bio-insecticides in controlling the *Ae. aegypti*  
95 | mosquito. Therefore, ~~it is important~~ the main objective of this study was to examine evaluate the  
96 | Insecticide Sscore of the toxin contained in the bioactive compounds of *P. paniculata* in killing  
97 | the *Ae. aegypti* mosquito.

## 98 | 2. Materials and Methods

### 101 | Preparation of Materials and Toolsequipment

102 | This study used 2500 ~~gramg~~ *P. paniculata*, 5 literre of 96% ethanol, 5 literre of distilled  
103 | water, ~~one~~ bottle of synthetic insecticide Baygon (cypermethrin), and 480 *Ae. aegypti*  
104 | ~~mosquito mosquitoes~~. The equipment used in this study ~~includesincluded~~ a blender, analytical  
105 | scales, Rotary Vacuum Evaporator, stopwatch, ~~shaker~~ water bath, thermometer, hygrometer,  
106 | Buchner funnel, stick, basin, test box, syringe, and a spray bottle.

Comment [Editor5]: Remark: Do you mean "a shaking water bath"?

### 108 | Test Animal Preparationanimal preparation

109 | Test animals were bred ~~usingin~~ media containing clean water in a cool place and protected  
110 | from direct sunlight for the *Ae. Aegypt#aegypti* mosquitoes to lay their eggs. The larvae were  
111 | reared in an aquarium at 24.2<sup>o</sup>C<sub>2</sub> - 24.4<sup>o</sup>C<sub>4</sub> <sup>o</sup>C, with a relative humidity of 67-70%. The larvae  
112 | ~~were fed were fed~~ with coconut water [28] ~~to become larvae until they reached~~ instar III and IV  
113 | stages before becoming mosquitoes. The adult *Ae. aegypti* mosquitoes were then used as test  
114 | animals.

### 115 | Extraction Proecessprocess

116 | A mass of 2500 ~~gramsg~~ of *P. paniculata* plants were washed and air-dried at room  
117 | temperature, and blended ~~them to form~~ give a final mass of 400 ~~gramsg~~ of powder. The powder  
118 | was macerated with 96% ethanol until completely submerged for three days. The resulting  
119 | solution was filtered using a Buchner funnel and placed in a dark bottle. ~~Furthermore, the~~ The  
120 | dregs from the first filtering were ~~then~~ soaked again for one day, filtered, and the process  
121 | repeated ~~infor~~ the third ~~immersion time~~. The ~~results~~ filtrates from the three maceration processes  
122 | were combined and concentrated using a Vacuum Rotary Evaporator to evaporate 96% ethanol  
123 | and obtain an extract. The extract obtained was stored in the refrigerator to be used later [29].

### 124 | Bio-insecticide's ExtractTests of extracts of *P. paniculata* Test against *Ae. Aegypti* 125 | Mosquitosmosquitoes

126 | Bio-insecticide's extractTests of *P. paniculata* ~~testsextracts~~ against *Ae. aegypti* ~~was carried~~  
127 | ~~outwere performed~~ by transferring 20 *Ae. aegypti* in each test ~~boxes, then sprayed~~ box before  
128 | spraying —with ~~various concentrations of~~ different concentrations of *P. paniculata* ~~plant~~  
129 | ~~extraextracts~~ with four repetitions per concentration, as well as for C (+) and C (-) K (+) and  
130 | K(-).The effect of the ~~concentrations~~ extracts on the *A. aegypti* mosquito was observed by  
131 | looking at the changes in behavior, movement, and physical condition until death. Deaths of Ae.  
132 |

Comment [Editor6]: Remark: It is not clear what these are. Please clarify.

133 | *Aegypti* mosquitoes ~~death~~ were counted every ~~five minutes~~ 5 min for one hour. Moreover, the  
134 | ~~in~~ insecticide ~~S~~ score of *P. paniculata* was determined from the number of *Ae. Aegypti* mosquitoes  
135 | considered ~~dead at~~ dead at 5-minute intervals. ~~Ae. Aegypti~~ *Aedes. aegypti* mosquitoes that  
136 | remained alive were left to die or killed with Baygon.

### 137 | Data analysis

138 | Data were analyzed using the ~~Statistical Test of One-Way~~ Analysis of Variance with RAL,  
139 | ~~followed by the One-Way ANOVA~~ test. However, when the ~~One-Way ANOVA test does~~ data did  
140 | not ~~meet conform~~ to the assumptions of parametric tests the requirements, ~~the non-parametric~~  
141 | Kruskal–Wallis ~~Non-Parametric Test test~~ and ~~the Test Spearman Analysis are~~ the Spearman test  
142 | ~~were~~ conducted to examine the relationship between the independent and dependent variables.

## 143 | 3. Results and Discussion

### 144 | A. Results

#### 147 | ~~The Results~~ Observations on deaths of ~~Observing Ae.aegypti Mosquitoes' Death After~~ 148 | ~~Spraying Mosquitoes after spraying the P. paniculata Plant Extract~~ plant extracts

149 | The *Ae. aegypti* mosquitoes died after ~~spraying the P.~~ exposure to concentration of 10% and 15  
150 | % ~~P. paniculata plant extract at 10% and 15% concentrations, extracts.~~ They died slowly by  
151 | flying irregularly and actively ~~and before~~ falling in a tilted body position. ~~The After falling, the~~  
152 | mosquitoes appeared weak, with ~~some legs still moving~~ limited leg movements, before becoming  
153 | paralyzed, ~~and eventually~~ dying, ~~and their bodies remaining stiff.~~ The death was faster at 20%  
154 | and 25% concentrations. In the positive control (K+), death occurred in less than five minutes,  
155 | while in the negative control (K-), ~~there was no death.~~

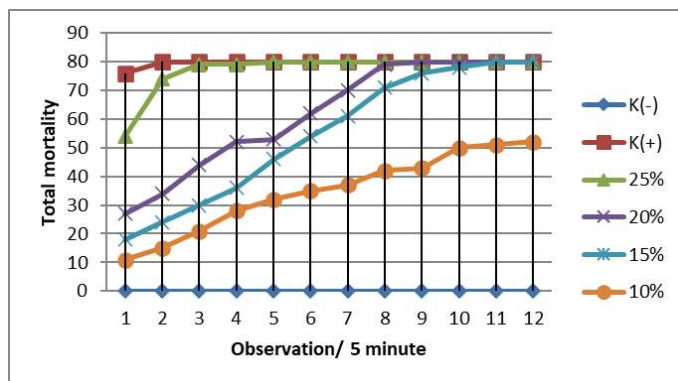
156 | ~~the Ae. Aegypti~~ mosquitoes tried to avoid the spray during the four experiment repetitions.

**Comment [Editor7]:** Remark: Check if this is correct. You later refer to deaths at 20% and 25%.

**Comment [Editor8]:** Remark: For a balanced comparison you should state whether death occurred or not in the negative control.

#### 161 | ~~Total mortality of Ae. aegypti Mosquitoes~~ Total Mortality mosquitoes at Each 162 | ~~Concentration With Four Repetition~~ each concentration

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164 | ~~Based on Figure 1 In the treatment group, the~~ After exposure to *P. paniculata* plant  
165 | extract ~~was sprayed with at~~ 10%, 15%, 20%, and 25% concentration, total mortalities of 11, 18,  
166 | 27 and 54 individuals, respectively, were recorded in the first 5 ~~minutes~~ min of observation-  
167 | (Figure 1). After 60 mins, the total mortalities of *Ae.aegypti* mosquitoes ~~death rate with 4~~  
168 | repetitions in a row was 11, 18, 27 and 54 individuals. In the 60 minutes, the total mortality of  
169 | *Ae. aegypti* mosquitoes in 4 repetitions ~~was were~~ 52, at 10%, 80, 80 each at 15%, 20%, and  
170 | ~~80-25%~~. In the positive control (K+) ~~using~~+, which was a synthetic insecticide baygon (~~called~~  
171 | cypermethrin) in less than 5 minutes, 76 *Ae. aegypti* mosquitoes fell and died in 4 repetitions,  
172 | namely 76 individuals. While in the less than 5 min (Figure 1). In the negative control (+) using  
173 | (K-), which was distilled water ~~did not cause death.~~ There was 80 *Ae. aegypti* mosquitoes were  
174 | ~~death~~ did not die. There were 80 *Ae. aegypti* mosquitoes deaths after 51–55 min at a  
175 | concentration of 15% ~~occurred at 51–55 minutes,~~ after 41–45 min at 20% concentration of 80  
176 | total deaths ~~occurred at 41–45 minutes~~ and a 25% concentration of 80 total deaths occurred  
177 | ~~after 21–25 minutes~~ at 25% concentration

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181 Figure 1: ~~Ae. Total mortality of Aegypti mosquitoes total mortality~~ at each concentration ~~with~~  
 182 ~~four repetitions at five-minute intervals~~

183 (Primary Data, 2020)

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185 The Average Mean mortalities of Ae. aegypti Death Rate at Each Concentration Every Five  
 186 Minutes Observation each concentration at five-minute intervals

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188 Based on Figure 2, the treatment group was sprayed with After exposure to *P. paniculata*  
 189 plant extract with a concentration at concentrations of 10%, 15%, 20%, and 25%, the mean  
 190 mortality of *Ae. aegypti* mosquitoes were 2.75 individuals (13.75%), 4.5 individuals (22.5%),  
 191 6.75 individuals (33.75%), and 13 individuals (67.5%), respectively, in the first 5 minutes in  
 192 observation, the average (Figure 2). Mean mortality rate of *Ae. aegypti* mosquitoes with 4  
 193 repetitions in a row was 2.75 tails (13.75%), 4.5 tails (22.5%), 6.75 tails (33.75%) and 13 tails  
 194 (67.5%). *Ae. aegypti* mosquitoes death rate in after 60 minutes with 4 repetitions at a concentration  
 195 of 10% was in of exposure were 13 tails individuals (65%), while the (%) at 10%, 20 individuals  
 196 (100%). at each of concentrations of 15%, 20%, and 25% were 20 tails (Figure 2). All (100%)  
 197 95%) *Ae. aegypti* mosquito individuals fell and died in positive control (K+) using synthetic  
 198 insecticide Baygon (cypermethrin) in less than 5 minutes with 4 repetitions. While min. In the  
 199 negative control (-) using distilled water did not cause death. *Ae. Aegypti* mosquitos totally (K-) no  
 200 deaths (0% mortality) were recorded. All (100%) of *Ae. aegypti* individuals died after 51-55 min  
 201 at 15% concentration in 51 until 55 minutes, after 41-45 min at 20% concentration of 80 total  
 202 deaths occurred at 41-45 minutes, and after 21-25 min at 25% concentration of 80 total deaths  
 203 occurred at 21-25 minutes. (Figure 2).

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Comment [Editor9]: Remark: 'Mean' is more academic than 'average'.

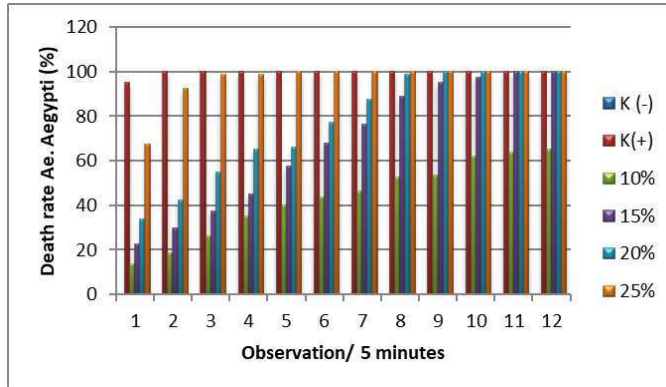


Figure 2: The Average Mortality Mean mortality of *Ae. aegypti* at each Concentration of Five Minutes Observation concentration at five-minute intervals

Based on these results of study in figure 1 and 2, explained indicate that the different concentrations of *P. paniculata* plant extracts gave had different effects on number of *Ae. aegypti* mosquitoes death in each treatment and repetition. The number of *Ae. aegypti* mosquitoes death tends mortality of *Ae. aegypti* individuals. Mortality tended to increase along with the increase in the concentration of *P. paniculata* plant extracts. It's means, indicating that the higher the concentration of extract used of the concentration, the higher of potency of the *P. paniculata* plant extract as a bio-insecticide against the *Ae. Aegypti* mosquito. During one hour observations showed the increasing number of *Ae. Aegypti* mosquitos's death. It's explained the longer an observation time, the greater potential as a bioinsecticide. *Aegypti*. This is was supported by the results of the Kruskal-Wallis test (p-value of = 0.001 < 0.05), which means that there is was a significant difference mortality rate of the *Ae. aegypti* mosquito between the death rate of the *Ae. aegypti* mosquito and the difference in the concentration of the concentrations of *P. paniculata* plant extract extracts (10%, 15%, 20%, 25 %) of Knockdown Time acceleration. The results of the Kruskal-Wallis test in this study were correct and continued with concurred with those of the Spearman correlation test with the results of p-value 0.008 < 0.05 there was which showed a significant correlation (p = 0.008) between the increase in the concentration of the extract (treatment group) extracts and the knockdown time of the *P. paniculata* plant extract. From the results of the Kruskal Wallis test and the Spearman correlation test, it can be concluded. These results indicate that the greater of higher the concentration, the faster shorter the time of to death of *Ae. aegypti* mosquito, the strength of the correlation is denoted by a correlation coefficient 0.312 (31.2%). Correlation coefficient occurred with positive value means that correlation is moderate. The higher concentrations of *P. paniculata* plants extract then the faster shorter of knockdown time. It was also noted that mortality increased with increased duration of exposure to the plant extracts

**Comment [Editor10]:** Remark: Please check this. I think there must be a negative correlation between concentration and time to death.



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**Insecticide Score of *P. paniculata* Plantsplant extracts**

Table 1 AverageMean falling down times of *Ae. Aegypti* Falling Down At Various Treatment Concentrations With*aegypti* at various treatment concentrations with Knockdown Time<sub>50</sub> (KT<sub>50</sub>)

Time	10%	15%	20%	25%	K (+)	K (-)
5	2,75	4,5	6,75	13,5	19	0
10	3,75	6	8,5	18,5	20	0
15	5,25	7,5	11	19,75	20	0
20	7	9	13	19,75	20	0
25	8	11,5	13,25	20	20	0
30	8,75	13,5	15,5	20	20	0
35	9,25	15,25	17,5	20	20	0
40	10,5	17,75	19,75	20	20	0
45	10,75	19	20	20	20	0
50	12,5	19,5	20	20	20	0
55	12,75	20	20	20	20	0
60	13	20	20	20	20	0

Legend:  $\square$ : *Ae. Aegypti* Knockdown Time (KT<sub>50</sub>);  
Source: (Primary Data, 2020)

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Knockdown Time<sub>50</sub> (KT<sub>50</sub>) is the time to knockdown of *Ae. aegypti* mosquitoes. In Table 1, the average repetition of the study show during the 10% concentration of *P. paniculata* plant extract had a The highest KT<sub>50</sub> of between 36 until and 40 minutes, 15 was recorded for the 10% concentration of *P. paniculata* plant extract followed by that of the 15% concentration had a KT<sub>50</sub> of between 21 until and 25 minutes, in min, and that of the *P. paniculata* plant extract with 20% concentration the KT<sub>50</sub> was obtained between 11 and 15 minutes min, - and in that of the *P. paniculata* plant extract with a 25% concentration of 25%, the KT<sub>50</sub> was less than 5 minutes. In the average iteration of the study, there in (Table 1). There were no *Ae. aegypti* mosquitoes that fell on died in the negative control and the Knockdown Time<sub>50</sub> (KT<sub>50</sub>) was less than 5 minutes in KT<sub>50</sub> of the positive control. This means was less than 5 min (Table 1). These results indicate that the different concentrations of *P. paniculata* plant extracts gave had different effects KT<sub>50</sub> on the number of *Ae. aegypti* that fell, as well as on each treatment and repetition mosquito.

Table 2: Bio-Insecticide Extracts scores of e of *P. paniculata* plant extracts based on Knockdown Time<sub>50</sub>

Group Control	KT50 (Time)	Knockdown Effect	Insecticide Score	Interpretation
Concentration 10%	36-40	-	1	-

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Concentration 15%	21–25	-	2	-
Concentration 20%	11–15	+	3	Weak Knockdown
Concentration 25%	< 5	+++	5	Quick Knockdown
Positive Control	< 5	+++	5	Quick Knockdown

Interpretation Data of data (+ weak Knockdown +++ Quick Knockdown) [30]  
(Primary Data, 2020)

Table 2 shows the effectiveness level of the four *P. paniculata* plant extract concentrations and positive control based on Insecticide Knockdown Time<sub>50</sub> (KT<sub>50</sub>). A 10% concentration of the *P. paniculata* plant extract had a KT<sub>50</sub> between 36 and 40 minutes. This implies min giving an Insecticide Score of 1 or no knockdown effect. Similarly, a 15% concentration of the plant extracts had a KT<sub>50</sub> of between 21 and 25 minutes, indicating an Insecticide Score of 2 or no knockdown. A 20% concentration of the plant extract had a KT<sub>50</sub> of between 11–15 minutes, implying min, giving an Insecticide Score of 3 or a weak knockdown effect. Furthermore, a 25% concentration of *P. paniculata* plant extract had a KT<sub>50</sub> in of less than 5 minutes. This indicates min, indicating an Insecticide Score of 5 or a quick knockdown effect. The 25% concentration had a KT<sub>50</sub> in of less than 5 minutes, similar to that of the positive control, with an Insecticide Score of 5 or a quick knockdown effect. Therefore, the 25% concentration of *P. paniculata* plant extract was the most effective in knocking out *Ae. Aegypti* mosquitoes, with an Insecticide Score of 5 or a quick knockdown effect.

## B. Discussion

The results in Figures 1 and 2 showed that the number and percentage of *Ae. aegypti* mosquito deaths increased with the concentration of *P. paniculata* plant extract. Higher This was expected as higher concentrations increase of the accumulation of P. paniculata plant extract's toxic extract a higher exposure of A. aegypti mosquitoes to higher levels of the bioactive compounds in A. aegypti mosquitoes' bodies, increasing their mortality.

Kosini examined the effect of the Gnidia kaussiana (Thymelaeaceae) extract on reported similar results for Callosobruchus maculatus. The study explained that larvae exposed to Gnidia kaussiana plant extracts and attributed them to an increased absorption of toxic compounds in by the larvae at higher concentration of the Gnidia kaussiana plant extract, which accelerates the mortality process of larvae Callosobruchus maculatus by melanizing the cuticle. This. Absorption of toxic plant bioactive compounds disrupts the endocrine system due to the presence of secondary metabolites such as terpenoids, alkaloids, and flavonoids [31]. The study of da Botas et al explained that essential oil attributed the laricide effect of Baccharis reticularia DC and limonene as a larvicide agent on Controlling Ae. Aegypti (Diptera: Culicidae), it's also able aegypti to its ability to inhibit the formation of acetylcholinesterase enzyme by blocking the nerve signal of transduction, which can cause the death and paralysis in Ae. aegypti larvae. Aegypti larvae. The more the larvae of Ae. aegypti absorb absorb the toxic compounds in B. reticularia essential oil, the more higher the Ae. aegypti death mortality rate.

**Comment [Editor11]:** Remark: It is discouraged to refer to figures in the Discussion.

299 Also, the longer the exposure to *B. reticularia* essential oil compounds ~~will increase~~ the higher  
300 the level of toxicity- [32].

301 ~~Many~~ Our results corroborate those of many previous studies ~~stated~~ stating that ~~the~~  
302 bioactive plant compounds have insecticidal, larvicidal, ~~repellent, and environmentally friendly~~  
303 effects ~~useful for insect control~~ insects. According to Suluvoy, essential oils, flavonoids,  
304 alkaloids, glycosides, esters, and fatty acids ~~have anti-insect effects~~. ~~Therefore, they could~~ can be  
305 used as an alternative to chemical compounds ~~in insect control as repellents, feeding deterrents~~  
306 ~~or antifeedants, toxicants, inhibitors, growth factors, chemosterilants, and attractants~~ because they  
307 have anti-insect effects [21].

308 Previous studies ~~explained~~ reported that *Polygala* plants contain bioactive compounds  
309 with various biological activities- such as alkaloids, saponins, flavonoids, phenols, tannins,  
310 steroids, and terpenoids [27]. ~~Tannins and flavonoids are phenolic plant compounds that act as~~  
311 ~~primary antioxidants or free radical scavengers~~ [33]. ~~The bioactive compounds of P. paniculata~~  
312 ~~able to affect the mortality of Ae.~~ The bioactive compounds in P. paniculata extracts able to  
313 cause mortality of Ae. -aegypti mosquitoes are flavonoids, saponins, tannins, alkaloids, steroids,  
314 and terpenoids. In this study, ~~the~~ mortality of Ae. aegypti mosquitoes ~~death~~ was most probably  
315 caused by compounds in the P. paniculata plant extract. ~~The compounds they~~ entered the  
316 mosquito's body through contact or respiratory poisoning and the mouth and digestive tract,  
317 causing stomach poisoning.

318 The ~~analysis shows that~~ results showed that the ~~number~~ mortality rate of *Ae. aegypti*  
319 mosquitoes ~~that died~~ when exposed to the *P. paniculata* plant extract varied with the ~~extract's~~  
320 concentration. ~~Variations in~~ of the ~~concentrations~~ affected the mortality of *Ae. aegypti*  
321 mosquitoes differently in each treatment and repetition. The results in Figures 1 and 2 show that  
322 ~~spraying a 25% concentration of the P. paniculata plant extract for 60 minutes kills 80 Ae.~~  
323 ~~Aegypti mosquitoes in less than the first five minutes.~~ This is the largest number of deaths that  
324 occurred faster than other concentrations. Therefore, the higher ~~the~~ concentration of the plant  
325 extract ~~increased its effectiveness~~ the more effective it was as a ~~vegetable~~ an insecticide against  
326 mosquitoes.

327 This study compared the effectiveness ~~level~~ of the four *P. paniculata* plant extract  
328 concentrations with ~~that of the~~ positive ~~controls~~ control based on Insecticide Knockdown Time<sub>50</sub>  
329 (KT<sub>50</sub>). A 25% concentration of the plant extract had a KT<sub>50</sub> of less than five minutes,  
330 ~~implying which was similar to that of the positive control.~~ This KT<sub>50</sub> implies an Insecticide Score  
331 of 5 or a quick knockdown effect. This is in line with the 2006 WHO standard, which stated that  
332 an insecticide ~~is~~ has a knockdown time required to drop a vector when the median knockdown  
333 ranges between 3-5 min [30]. Furthermore, ~~it an insecticide~~ has a quick knockdown effect when  
334 it has a KT<sub>50</sub> of less than ~~five minutes~~ 5 min. According to Norris, a good insecticide  
335 requirement for controlling disease vector insect species ~~is that it~~ must cause a rapid knockdown  
336 of the target species, ~~especially active pathogens.~~ Additionally, it must quickly intervene and kill  
337 adult mosquitoes [17].

338 The 25% concentration had a KT<sub>50</sub> of less than five minutes, similar to a positive control  
339 ~~Baygon containing cypermethrin.~~ They both had an Insecticide Score 5, ~~implying a quick~~  
340 ~~knockdown effect.~~ Therefore, the 25% concentration of the *P. paniculata* plant extract was the  
341 most effective concentration in knocking down *Ae. aegypti* mosquitoes. ~~This is because as~~ it had  
342 an Insecticide Score of 5 or a quick knockdown effect. The positive control treatment was  
343 ~~intended to compare a benchmark for comparing~~ the quality of *P. paniculata* plant ~~extract~~  
344 ~~concentration~~ extracts. In contrast, the negative control treatment was used to compare its

Comment [Editor12]: Remark: This is not relevant to your study. Hence, we have deleted this.

Comment [Editor13]: Remark: We have deleted this as it is a repetition of the results.

345 effectiveness with that of the plant extract. ~~The results showed that no mosquito died after 60~~  
346 ~~minutes of observation.~~

347 Chang highlighted the need to use bio-insecticides as an alternative insect controller.  
348 ~~Using inappropriate insecticides causes insects to adapt easily by metabolic detoxification~~  
349 ~~quickly and survival. This causes synchronization of insect immunity passed on to their~~  
350 ~~offspring. Therefore, it~~ is necessary to develop safe alternative insecticides, larvicides, and  
351 repellents effective for humans, animals, the environment, and the ecosystem. Natural  
352 insecticides are needed to suppress vector resistance and slow down their genetic adaptation [34].  
353 According to Sulovoy and Grace, botanical plant insecticides only affect target insects, do not  
354 destroy beneficial natural enemies, and are a safe and residue-free food source. [21]. It is more  
355 environmentally friendly, effective, cheap, and naturally available [12].

**Comment [Editor14]:** Remark: Reference 33 has not been cited in the manuscript. Please check and revise.

**Comment [Editor15]:** Remark: There is no clear link of this paragraph to your results. Please check and revise.

## 356 Conclusion

357 The ~~active substance~~ bioactive compounds contained in the *P. paniculata* plant ~~extract~~  
358 ~~has extracts~~ have the ability to drop and kill *Ae. aegypti* mosquitoes. ~~This is because more~~  
359 ~~compounds in the plant extracts exposed to the mosquitoes increase the knockdown effect time.~~  
360 Therefore, the plant ~~extract~~ extracts *P. paniculata* are a potential alternative insecticide for  
361 controlling *Ae. aegypti* mosquitoes. ~~It does~~ Like other plant insecticides, it is expected that *P.*  
362 *paniculata* extracts will not leave residues in the environment and ~~is safe for other living~~  
363 ~~beings will be environmentally friendly.~~ Also, the ~~extract~~ bioactive compounds ~~dein the~~  
364 extracts will not cause resistance against *Ae. aegypti* mosquitoes, ~~and the plant has economic~~  
365 ~~value and is beneficial to cultivate.~~

## 366 Acknowledgments

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368 with the title "Bio-insecticides Extract of Scented Root (*Polygala paniculata*) in Controlling  
369 *Aedes aegypti* mosquito" with Contract Number No. 08/STIKes-HTP/VI/2020/0147, A.

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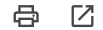
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